

LEAD BIOACCUMULATION IN WHOLE BODY TISSUES OF FRESH WATER GASTROPOD SNAIL, *BELLAMYA BENGALENSIS* AFTER CHRONIC LEAD INTOXICATION

***Mahajan P.R.**

Department of Zoology, Sardar Vallabhabhai Patel Arts and Science College, Ainpur, Tal - Raver, Dist – Jalgaon. 425509

**Author for Correspondence*

ABSTRACT

The objective of present study was carried out to determine the level of bioaccumulation of heavy metal salt lead (Pb) in freshwater gastropods snail's *Bellamya bengalensis*. The bioaccumulation of heavy metal salts in snail's was studied under two groups. Group A was maintained as control, group B snail's were exposed to chronic LC_{50/10} dose of lead nitrate (6.753 ppm) for 21 days. Bioaccumulation level in whole body tissues of *Bellamya bengalensis* from A and B groups were collected after every seven days and were dried at 80^oc in an oven till constant weight was obtained. The sample were analysed on the instrument atomic absorption spectrophotometer (Chemito). It was found that the freshwater snail, *Bellamya bengalensis* showed the highest concentration of lead as compared to control, lead contamination of the aquatic ecosystems affect the life of the snails, altering their metabolic functions. Hence, a scientific detoxification method is essential to improve the health of economic species of snail's in any stressed environmental conditions (accidental or induced discharges of heavy metal).

Keywords: Chronic Lead Intoxication, Bioaccumulation, Fresh Water Snail

INTRODUCTION

In aquatic ecosystem, heavy metals are considered as the most important pollutants, since they are present throughout the ecosystem and are detectable in critical amounts. Generally, metals enter the fresh water bodies from a variety of sources, including: rocks and soils directly exposed to waters, dead and decomposing vegetation and animal matter, wet and dry fallout of atmospheric particulate matter and human activities, including the discharge of various treated and untreated wastes to the water body (Abo *et al.*, 2005). Non-essential metals are usually potent toxins and their bioaccumulation in tissues lead to intoxication, decreased fertility, tissue damage and dysfunction of a variety of organs (Oliveira *et al.*, 2000; Damek-Proprawa and Sawicka-Kapusta, 2003) Balkas (1982) reported that, heavy metal pollution of terrestrial and aquatic ecosystems have long been recognized as a serious environmental concern. This is largely due to their non-biodegradability and tendency to accumulate in plants and animals tissues. As a result, metal bioaccumulation is a major route through which increased levels of the pollutants are transferred across food chains/web, creating public health problems wherever man is involved in the food chain (Tuzen, 2003; Otitoloju and Don-Pedro, 2002, 2004).

Lead (pb) is a heavy metal which occurs naturally or result from industrial contamination, or be leached from lead pipes in some water systems. Lead is widely used in paint industry, pigments, dyes, electrical components, plastic chemicals and in various other industries (Hodson *et al.*, 1984). Since some of the Pb salts are soluble in water, presents a potential threat to aquatic organisms. However enrichment of Pb within an aquatic organisms and it's transfer through food chain constitute a danger to man. The strong attraction between metal ions and organic ligands influence the deposition of metals in the body and their rate of excretion, consequently, with continuing intake, it tends to accumulate to a high degree in the body of molluse (Harrison, 1969; Simkiss and Mason, 1983). The bioaccumulation of heavy metals in the different fish tissues has been studied by several investigators (Filazi *et al.*, 2003 and Ashraf, 2005). Owing to the gastropods' potential characteristics as biomonitors of heavy metal pollution, this study focused on snail, *Bellamya bengalensis*. The objective of the present study was to determine the heavy metal concentrations of Pb in whole body.

Research Article

MATERIALS AND METHODS

The snail, *bellamya bengalensis* were acclimatized to laboratory condition for 2-3 days and healthy active snail's of approximately medium size and weight were chosen. These snail's were divided into two groups, such as group A and B. The snail's of group A were maintained as control. The snail's from group B were exposed to chronic concentration (LC 50 value of 96 hr/10) of heavy metal salt, lead nirtate (6.753 ppm) upto 21 days. During experimentation snails were fed on fresh water algae. The whole body mass of snails from all groups were collected after every seven days and were dried at 80 c in an oven till constant weight was obtained. The 500 mg sample was taken for digestion. The tissue was digested in 10 ml of acid mixture (HCL:HNO₃ in (3:1) ratio) on hot plate till dryness. The digested mixtures were kept in water bath for 6-7 hours until the samples were cooled. Cool digested samples were filtered (Whatman grade 541). The total volume was diluted to 50 ml by double glass distilled water in volumetric flask. The sample were analysed on the instrument atomic absorption sepectrophotometer (Chemito). The concentration of Pb accumulation in the tissue of each exposure period was recorded and the results are given in the table.

RESULTS AND DISCUSSION

Observation and Results

Lead contents in *Bellamya bengalensis* after exposure to concentrations of lead nitrate (6.753 ppm) up to 21days. After 7, 14 and 21 days of chronic exposure to heavy metal, it was observed that there was an increase in concentration of accumulated heavy metals in the body of *B. bengalensis* with respect to time as compared to those of control snails.

The accumulation data from table and Figure indicates that the concentration of bioaccumulated lead in presence of PbNO₃ (6.753ppm) increased with increase in exposure period as compared to control. The lead content is expressed in µgm/kg. dry weight. The control group of animals showed minute quantity of lead as compared to the experimental groups. The control group of animal showed 730.0 µgm/kg, lead in whole body tissue while the bioaccumulated lead in presence of PbNO₃ (6.753 ppm) after 7 days exposure was 1792.0µgm/kg. The concentration in the tissues was raised after 14 days to 2652.0µgm/kg, while after 21 days increases to 3646.0µgm/kg. There was more change in the bioaccumulation of Pb in control animals.

Table 1: Lead content (µgm/kg dry weight) in whole body of *bellamya bengalensis* (lamarck) after chronic treatment of lead nitrate

Treatment	Sr No.	Body Tissue	Hg content (µgm/kg dry weight)		
			7 Days	14 Days	21 Days
(A) Control	i	W.B.	730.00	730.00	724.00
(B) 6.753 ppm PbNO ₃	ii	W.B.	1792.00 + 59.263*	2652.00 + 72.473*	3646.00 + 71.714*

W.B.- Whole Body / • -Compared with respective A

Discussion

In the present study, the freshwater gastropod snail's *Bellamya bengalensis* were exposed to LC50/10 concentrations of for twenty one days in the laboratory to examine the effects of the heavy metals on their survival and their fate through their soft parts. From the obtained results it is clear that the analysis of the investigated gastropods snail's (whole body tissue's) indicated that these organisms can accumulate Pb in high concentrations in their whole bodies, so they can be used as bioindicators for heavy metals pollution in aquatic ecosystems. Bakre and Garg (1994) studied bioaccumulation of Pb in experimentally exposed freshwater mollusc, *Pila globosa* and maximum accumulation was found in Intestive, followed by digestive gland and less degree of accumulation of Pb occurred in mantle and foot. The accumulation of Pb increased with increasing exposure period. According to the Gundacker (1999), a zebra mussel

Research Article

accumulates high amounts of potentially toxic metals and was widely used as a bio-monitoring organism. Avelar *et al.*, (2000) reported that Oyster and mussels can accumulate Cd in their tissues at levels up to 100,000 times higher than the levels observed in the water in which they live. Passow *et al.*, (1961) reported that lead can induce synthesis of specific proteins which selectively bind them. Inhibition of enzyme activities by heavy metals is either due to the direct binding with enzyme protein or due to damage of cell organelles or by toxic effect produced. The specific amoebocytes and or digestive vesicles within the cell may engulf metals outside the cell membrane (i.e. in the human digestive tract), then move back into the tissue carrying their particulate burden (Owne *et al.*, 1966).

ACKNOWLEDGEMENT

The authors are thankful to the Principal, Dhanaji Nana Mahavidyalaya, Faizpur for providing the laboratory facility to carryout the work.

REFERENCES

- Abo El Ella SM, Hosny MM and Bakry M (2005).** Utilizing fish and aquatic weeds infestation as bioindicators for water pollution in Lake Nubia, Sudan. *Egyptian Journal of Aquatic Biology and Fisheries* **9** 63-84.
- Ashraf W (2005).** Accumulation of heavy metals in kidney and heart tissues of *Epinephelus microdon* fish from the Arabian Gulf. *Environmental Monitoring and Assessment* **101** 311-316.
- Avelar WEP, Mantelatto FLM, Tomazelli AC, Silva DML, Shuhama T and Lopes JLC (2000).** The marine mussel *Perna perna* (Mollusca, Bivalvia, Mytilidae) as an indicator of contamination by heavy metals in the Ubatuba Bay, Sao Paulo, Brazil. *Water, Air and Soil Pollution* **118** 65-72.
- Bakre PP and Garg S (1994):** Bio-accumulation of lead in experimentally exposed freshwater mollusc, *Pila globosa*. *Indian Journal of Environment and Toxicology* **4**(2) 19 -24.
- Balkas IT, Tugrel S and Salhoglu I (1982).** Trace metals levels in fish and crustacean from Northeastern Mediterranean coastal waters. *Marine Environmental Research* **6** 281-9.
- Damek-Proprawa M and Sawicka-Kapusta K (2003).** Damage to the liver, kidney and testis with reference to burden of heavy metals in yellownecked mice from areas around steel works and zinc smelters in Poland. *Toxicology* **186** 1-10.
- Filazi A, Baskaya R, Kum C and Hismiogullari SE (2003).** Metal concentrations in tissues of the Black Sea fish *Mugil auratus* from Sinop-Icliman; Turkey. *Human and Experimental Toxicology* **22** 85-87.
- Gundacker C (1999).** Tissue specific heavy metals (Cd, Pb, Cu and Zn) deposition in Natural pollution of Zebra mussel (*Dreissena polymorpha* Pallas). *Chemosphere* **38** 3339-3356.
- Harrison RL (1969).** Accumulation and distribution of ⁵⁴Mn and ⁶⁵Zn in freshwater clams. In: *Radioecology*, edited by Nelson DJ and Evans FC (Conference 670503 U.S. Dept. Commer. Springfield, Virginia) 198-220.
- Hodson PV, Whittle DM, Wong PTS, Borgmann U, Thomas RL, Chau YK, Nriagu JO and Hallet DJ (1984).** *Lead Contamination Of The Great Lakes And It's Potential Effects On Aquatic Biota. Offprints From Toxic Contaminants In The Great Lakes*, edited by Nriagu JO and Simons MS (John Wiley and Sons. Inc).
- Oliveira Ribeiro, Pelletier CE, Pfeiffer W and Rouleau C (2000).** Comparative uptake, bioaccumulation and gill damages of inorganic mercury in tropical and nordic freshwater fish. *Environmental Research* **83** 286-292.
- Otitoloju AA and Don-Pedro KN (2002).** Bioaccumulation of heavy metals (Zn, Pb, Cu and Cd) by *Tympanotonus fuscatus* var. *radula* exposed to sublethal concentrations of the test metal compounds in laboratory assays. *West African Journal of Applied Ecology* **3** 17-29.
- Otitoloju AA and Don-Pedro KN (2004).** Integrated laboratory and field assessments of heavy metals accumulation in edible periwinkle, *Tympanotonus fuscatus* var *radula* (L.). *Ecotoxicology and Environmental Safety* **57**(3) 354-62.

Research Article

Owen G (1966). In: *Physiology of Mollusca II* edited by Wilbur KM and Younge CM (Academic Press) New York 53-88.

Passow H, Rothstein A and Clarkson TW (1961). The general pharmacology of the heavy metals. *Pharmacological Reviews* **13** 185.

Simkiss K and Mason AZ (1983). Metal ions: Metabolic and toxic effects. In: *The Mollusca, Volume 2: Environmental Biochemistry and Physiology*, edited by Hochachka PW and Wilbur KM (Academic Press) New York 164.

Tuzen M (2003). Determination of heavy metals in fish samples of the middle Black Sea (Turkey) by graphite furnace atomic absorption spectrometry. *Food Chemistry* **80** 119-23.