PROPAGATION OF *BACOPA MONNIERI* (BRAHMI): IMPORTANT MEDICINAL PLANT

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ABSTRACT

An experiments were conducted for standardization of *in-vitro* propagation technique of *Bacopa monnieri* (L.), a medicinally herbs of India. Bacopa monnieri (Brahmi) has been used in the various ancient traditional system of medicine for centuries including Ayurveda. Brahmioriginate naturally in India, has a long history of use in number of disorders particularly anxiety, intellect and poor memory and used as anti-inflammatory, anticancer and antioxidant activities, analgesic, antipyretic. Quality planting material is major factor for any economically important medicinal plants and Micro-propagation technique may play an important role. Healthy nodal segments of the herb were used as explants with basic MS medium for shoot initiation and multiplication containing various combinations of different growth regulators. For the initiation of ex-plant BA (0.1 to 0.6 mg/l), IA (0.1 to 0.5 mg/l) and NA (0.1 and 0.5 mg/l) was used, while concentration of BA (0.5 to 1.5 mg/l), IA (0.5 mg/l) and NA (0.5 mg/l) was used for multiplication. MS full and 1/2MS were used for rooting of plantlets with 25 to 150 mg/l Activated Charcoal (AC). Maximum mean number of initiated plantlets 1.8 ± 0.42 with mean length 3.00 ± 0.94 were found in MS medium treated with lowest concentration of BA, while maximum mean number of multiplied plantlets 10.00 ± 2.58 with mean length 6.1 ± 1.91 were found in MS medium treated with 1mg/1BA and 0.5mg/1 IA. Maximum mean number of roods 12.4 ± 1.074968 with Mean length 9.19 ± 0.68 was observed on 1/2MS medium with 100 mg/l activated charcoal. The rooted plantlets were successfully hardened in 1:1:1 ratio of sand: soil: vermicompost and successfully established in soil.

Keywords: Brahmi, Bacopamonniera, Micropropagation, Medicinal Plants, Plant Tissue Culture

INTRODUCTION

Brahmi is considered as the main rejuvenating herb, which played a very important role in Ayurvedic therapies (Kapil and Sharma, 2014). "Brahmi" has been used in the Ayurvedic system of medicine for centuries. Bacopa monnieri (L.), commonly known as "Brahmi", is a member of the Family Scrophulariaceae, is placed second in the priority list of Indian medicinal plants (Anonymous, 1997). It is commonly found on the banks of rivers and lakes. It has been used for centuries in legends and traditional system of medicine as a memory enhancer (Bhattacharya et al., 1998; Shankar and Singh, 2000), antiinflammatory (Williams et al., 2014), analgesic, Antidiarrhoeal, Cytotoxic activity (Afjalus et al., 2012), antipyretic (Bammidi et al., 2011), sedative and anti-epileptic agent (Srivastava et al., 2009). In addition to its unique medicinal use, Bacopa monnieri has also been linked to phytoremediation programmes for the removal of heavy metals such as cadmium and chromium (Subashri and Pillai, 2014). Brahmi leaves are oblong, sessile and fleshy. In the Ayurvedic Bacopa has been recognized for its brain enhancement personality. Bacopa monnieri is a small, creeping herb with numerous branches, small oblong leaves and light purple flowers. In India and the tropics it grows naturally in wet soil, shallow water and marches of Bacopa monnieri (Brahmi) (Mohapatra and Rath, 2005). Family Scrophulariaceae is also known as Madhyarasayana in Ayurveda as it increase mental-clarity and brain stimulating action, it also possesses anti-inflammatory, analgesic, antipyretic, epilepsy, insanity, anticancer and antioxidant activities (Satvavati et al., 1976; Jain et al., 1994; Elangovan et al., 1995; Tripathi et al., 1996; Vohora et al., 1997). Saponins such as bacosides A, B, C and D which are the active triterpenoid principles and known as "memory chemicals" (Rastogi et al., 1994). Only a very limited research has been carried out on the plant, under the present study assumes singular significance and it is supposed to CIBTech Journal of Biotechnology ISSN: 2319–3859 (Online) An Open Access, Online International Journal Available at http://www.cibtech.org/cjb.htm 2016 Vol. 5 (3) July-September, pp.17-23/Vijay et al.

Research Article

contribute a great deal to the existing literature. The micro propagation protocol of the medicinally important plant. *Bacopa monnieri* was standardized using nodal segments as explants. This review highlights mainly the recent development and achievements made for the multiple shoots and root regeneration of *Bacopa monnieri* (Brahmi). The present paper highlights the morphogenic response of various auxins and cytokinins on *Bacopamonniera*.

MATERIALS AND METHODS

Collection of Explants

Branches of Brahmi were collected from healthy growing plants from medicinal garden of Prof. T.S. Murthi Science and Technology Station Obedullagani, Raisen (M.P.)

Sterilization of Explants

Nodal explants were cut and washed in running tap water to remove the superficial dust particles and mud adhering to its surface. Explants were washed with sevelon (5-10 drops/100ml) in a vial by gentle agitating conditions. The explants were thoroughly rinsed with distilled water for several times. Again these explants were dipped in to the 1% fungicide (Bavistin) treatment was given for 15 minutes and then washed with distilled water. For surface sterilization, Explants were transferred to sterile empty flasks under aseptic conditions and given a quick dip in 70% alcohol and subsequently they were washed in distilled water. After that, the explants were surface sterilized with different concentration of sterile (HgCl₂) for different duration as per the treatment to find out the best treatment for sterilization of explants. To remove the traces of sterile explants were washed in sterilized distilled water at least 5-6 times. The procedure was carried out in the inoculation chamber under laminar air flow hood.

Preparation of MS Medium

Culture media was prepared as per described method of Murashig and Skoog (1962) and different growth regulator was added as per requirement. For the initiation of ex-plant BA (0.1 to 0.6 mg/l), IA (0.1 to 0.5 mg/l) and NA (0.1 and 0.5 mg/l) was used, while concentration of BA (0.5 to 1.5 mg/l), IA (0.5 mg/l) and NA (0.5 mg/l) was used for multiplication. MS full and 1/2MS were used for rooting of plantlets with 25 to 150 mg/l Activated Charcoal (AC) combination adding 30 g/l sucrose and 5.7% agar. The hormones used for experiment were taken from stock solutions, which were previously prepared and kept under cold condition in refrigerator (Doods and Roberts, 1985). The pH of the medium was adjusted to 5.7 with 0.1 NaOH before autoclaving at 15 lbs and 121°C for 18 min.

Aseptic Inoculations of Explants

Nodal segments about 0.5-0.8 cm were prepared aseptically and were implanted vertically on Surface disinfected nodal explants were inoculated onto full strength MS medium (Murashige and Skoog, 1962) fortified with specific concentrations of growth regulators. The cultures were incubated at a constant temperature of $26\pm2^{\circ}$ C with 16 ± 1 h photoperiod (3000 lux).

RESULTS AND DISCUSSION

Results

Surface Sterilization and Induction of Axillary Shoots

Treatment of explants with 0.1% HgCl2 for 3 minutes resulted 100% contamination-free viable cultures. Final observation after 3-4 weeks showed that MS media supplemented with 0.1 mg/l of BAP proved to be most capability in shoot induction. On this medium an average of 1.8 \pm 0.42 shoots with mean shoot length 3 \pm 0.94 cm were obtained (Table 1, Figure 1A).

Shoot Multiplication

Shoot multiplication is depend on different type of concentration. Sometimes BAP increasing is best for shoot or just opposite. Activated auxiliary shoots from the nodal explants and transfer to fresh medium containing 1.0 mg/l BAP and 0.5 IAA to establish a stock of shoots used for *in vitro* multiplication. When we look Results in the present study showed the essential of plant growth regulators for *in vitro* multiplication, as the shoots cultured on basal medium did not multiply and become dead. BAP and IAA at a concentration of 1.0 mg/l and 0.5 just gave an average of 10.00 ± 2.58 shoots with mean shoot length

 6.01 ± 1.91 cm after 3-4 weeks of culture (Figure 1B). Increasing the concentration of BAP to 3.0 mg/l, a Decrease in shoot multiplication rate was observed. Trials on effect of cytokinin: auxin ration on *in vitro* shoot multiplication of Brahmi showed that results were better than in medium with 3.0 mg/l BAP. However, comparative number, length and health of shoots on media with BAP + IAA/NAA were not as good as in media containing 1.0 mg/l BAP and 0.5 IAA (Table 2).

Table 1: Effect of Plant Growth Regulators on in Vitro Axillary Shoot Induction in B. Monnieri

S. No	MS + PGR (mg/l)			Observations after 3-4 Weeks		
	BAP	IAA	NAA	Mean Shoot Number	Mean Shoot Length (cm)	
Control	0.0	0.0	0.0			
1	0.1	0.0	0.0	1.8 ± 0.42	3.00 ± 0.94	
2	0.2	0.0	0.1	0.9 ± 0.99	1.7 ± 1.1	
3	0.3	0.5	0.0	0.7 ± 0.70	1.4 ± 1.17	
4	0.4	0.0	0.1	0.5 ± 0.70	1.6 ± 1.26	
5	0.5	0.0	0.0	0.4 ± 0.69	2.8 ± 1.13	
6	0.6	0.5	0.5	0.4 ± 0.51	1.3 ± 1.41	

Note: Each treatment consisted of 10 replications. Data (Mean ±SE) were recorded after 20 days of culture.

Table 2: Effect of Plant Growth Regulators on in Vitro Axillary Shoot Multiplication in B. Monnieri

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S. No	MS + PGR (mg/l)			Observations after 3-4 Weeks		
	BAP	IAA	NAA	Mean Shoot Number	Mean Shoot (cm)	Length
Control	0.0	0.0	0.0		0.00 ± 0.00	
1	0.5	0.5	0.0	3.5 ± 2.58	3.6 ± 1.89	
2	0.1	0.5	0.0	10.00 ± 2.58	6.1 ± 1.91	
3	1.5	0.0	0.5	4.7 ± 2.54	3.1 ± 1.52	
4	1.5	0.5	0.0	4.8 ± 2.61	3.22 ± 1.76	
5	2.0	0.0	0.0	4.4 ± 1.50	4.00 ± 2.05	

Note: Each treatment consisted of 10 replications. Data (Mean ±SE) were recorded after 20 days of culture.

In vitro Rooting

After 30 Days of growth, rooting growth is rarely increase day by day in best culture. The multiple-shoot clumps produced on this medium were transferred to solidified MS growth-regulator-free medium for shoot elongation and rooting. On opposite, shoots were also observed for rooting on full- or half-strength MS medium with Activated Charcol ensuing excellent response for root induction. Maximum rooting was recorded in medium containing 100 mg/l Activated Charcol Supplemented with $\frac{1}{2}$ MS medium (Figure 1C). On this medium an average of 12.4 ± 1.074968 roots with average root length 9.19 ± 0.6806 cm was observed after 3-4 weeks (Table 3).

Hardening

After 5 weeks complete rooting medium, they were transfer to pots soil: sand: manure (1:1:1) and maintained in greenhouse or poly house. In poly house is carefully monitored for growth, development and water done as must required. Long and healthy plants transferred to the field condition open ground where 100% growth rate was measured.



Figure 1A: Initiation of Bacopa Monnieri



Figure 1B: Multiplication of Bacopa Monnieri



Figure 1C: Rooting of Shoot Bacopa Monnieri

Discussion

Bacopa monnieri is a medicinal plant used in Ayurvedic medicine for thousands of years to treatment for mental illness, asthma, anxiety, and age-related, Antioxidant, Stress, cough and cold etc. Mercuric chloride is highly antimicrobial against for both fungi and bacteria at low concentrations (upto 0.1 %) it is perhaps the most effective disinfective agent for soil-borne fungi (Nwokocha *et al.*, 2015). In this study, 100% contamination-free viable cultures were obtained by treatment of explants with 0.1% HgCl2 for 3

minutes. From the present studies MS media proved to be the best culture medium for the establishment of shoot culture in B. monnieri plant. This work was undertaken in order to learn the tissue culture technique for medicinally important plant Bacopa which is the mostly used for memory enhancing purposes. Final observation after 3-4 weeks showed that MS media supplemented with 0.1 mg/l of BAP proved to be most capability in shoot induction. On this medium an average of 1.8 ± 0.42 shoots with mean shoot length 3 ± 0.94 cm were obtained. Numerous reports of BAP as bud inducer at concentrations ranging from 1.0-5.0 mg/l have already published (Shrivastava $et\ al.$, 1999; Binita $et\ al.$, 2005; Ramesh $et\ al.$, 2006; Sharma $et\ al.$, 2010; Chandra $et\ al.$, 2012; Kaur $et\ al.$, 2013; Mohanta $et\ al.$, 2014; Behera $et\ al.$, 2015).

These present results are supported by the findings of other workers who have also observed and experimentally found the positive influence of MS medium for optimum shoot and root multiplication in different Bacopa species. Activated axillary shoots from the nodal explants and transfer to fresh medium containing 1.0 mg/l BAP and 0.5 IAA to establish a stock of shoots used for *in vitro* multiplication. This observation is supported by previous studies on *B. monnierri* (Chaplot *et al.*, 2005; Ceasar *et al.*, 2009; Vijyakumar *et al.*, 2010; Kumari *et al.*, 2010; Showkat *et al.*, 2010; Yusuf *et al.*, 2011; Mehta *et al.*, 2012; Pandiyan and Selvaraj, 2012; Jain *et al.*, 2013; Asha *et al.*, 2013; Tanveer *et al.*, 2010; Kaur *et al.*, 2013; Behera *et al.*, 2015). MS medium with Activated Charcol ensuing excellent response for root induction. Maximum rooting was recorded in medium containing 100 mg/l Activated Charcol Supplemented with ½ MS medium. Tissue culture raised plants are need acclimatization before field transfer. For this purpose *in-vitro* regenerated plantlets were shifted to pots and kept in Polyhouse for about a month. The protocol resulted in development of healthy plants without any need of intermediary hardening treatment. After 5 weeks complete rooting medium, they were transfer to pots soil: sand: manure (1:1:1) and maintained in greenhouse or polyhouse. So, this technology is effective as it produces thousands of plants in a short span of time.

Table 3: Rooting Response of in Vitro Regeneration Excised Shoots (10 Repeats)

S. No.	Medium	Activated Charcoal (mg/l)	No. of Roots/ Plantlets (Mean_SE)
1	MS	-	6.6 ± 1.264911
2	$^{1/2}$ MS	-	6.9 ± 0.994429
3	$^{1/2}$ MS	25	7.7 ± 0.823273
4	$^{1/2}$ MS	50	9.2 ± 1.032796
5	$^{1/2}$ MS	100	12.4 ± 1.074968
6	$^{1/2}MS$	125	12.3 ± 1.251666
7	$^{1/2}$ MS	150	9.3 ± 1.766981

Note: Each treatment consisted of 10 replications. Data (Mean \pm SE) were recorded after 20 days of culture.

Conclusion

Bacopa monnieri has always been a topic of interest of researchers. From tissue culture point of view several studies have been performed to propagate the plant *in vitro*. The highlights of the study are Use of low concentration of plant growth regulators and minimization of time required for field transfer of tissue culture raised plantlets. Free plants produced open the scope for utilization of plant material for antimicrobial testing and suitable pharmaceutical preparations. Apart from this in vitro propagation of Bacopa monnieri showed a highest rate of multiplication which cannot be seen in naturally found species of Bacopa monnieri. The Bacoppa research will give a new insight of research in medicinal components of plants through various advance techniques.

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Research Article

REFERENCES

Afjalus SM, Chakma N, Rahman M, Salahuddin M and Kumar SS (2012). Assessment of Analgesic, Antidiarrhoeal and Cytotoxic activity of ethanolic extract of the whole plant of *Bacopa monnieri* linn. *International Research Journal of Pharmacy* **3**(10) 98-101.

Anonymous (1997). *Indian Medicinal Plants: A Sector Study*, Occasional paper No. 54. Export-Import Bank of India. (Quest Publication, Bombay, India).

Asha KI, Devi AI, Dwivedi NK and Nair RA (2013). In vitro regeneration of Brahmi (*Bacopa monnieri* (Linn.) Pennell - an important medicinal herb through nodal segment culture. *Research in Plant Biology* **3**(1) 01-07.

Bammidi SR, Volluri SS, Chippada SC, Avanigadda S and Vangalapati MA (2011). Review on pharmacological studies of Bacopamonniera. *Journal of Chemical, Biological and Physical Sciences* 1(2) Sec B 250-259.

Behera S, Nayak N, Shasmita, Barik DP and Naik SK (2015). An efficient micropropagation protocol of *Bacopa monnieri* (L.) Pennell through two-stage culture of nodal segments and ex vitro acclimatization. *Journal of Applied Biology & Biotechnology* **3**(3) 016-021.

Bhattacharya SK and Ghosal S (1998). Anxiolytic activity of a standardized extract of Bacopamonniera: an experimental study. *Phytomed* **5** 77-82.

Binita BC, Ashok MD and Yogesh TJ (2005). *Bacopa monnieri* (L.) Pennell: a rapid, efficient and cost effective micropropagation. *Plant Tissue Culture and Biotechnology* **15**(2) 167-175.

Ceasar SA, Maxwell SL, Prasad KB, Karthigan M and Ignacimuthu S (2010). Highly efficient shoot regeneration of *Bacopa monnieri* (L.) using a two-stage culture procedure and assessment of genetic integrity of micro propagated plants by RAPD. *Acta Physiologiae Plantarum* 32 443 452.

Chandra G, Kumar V, Mukhija S, Dhingra A, Rajpurohit S and Narula P (2012). *In vitro* regeneration of brahmi (*bacopamonneiri* (l.) Penn.) - a threatened medicinal plant. *Kathmandu University Journal of Science, Engineering and Technology* **8**(1) 97- 99.

Chaplot B, Dave A and Jasrai Y (2005). *Bacopa monnieri* (L.) Pennell: A Rapid, Efficient and Cost Effective Micropropagation. *Plant Tissue Culture and Biotechnology* **15**(2) 167-175.

Doods JH and Roberts W (1985). Experiments in Plant Tissue Culture, second edition, (Cambridge University Press, Cambridge, UK).

Dubey RC (1999). A Text Book of Biotechnology, (Publication, S. Chand and Co., New Delhi, India).

Elangovan V, Govindasamy S, Ramamoorthy N and Balasubramanian K (1995). *In vitro* studies on the anticancer activity of *Bacopa monnieri*. *Fitoterapia* 66 211–215.

Jain R, Prasad B and Jain M (2013). In vitro regeneration of *Bacopa monnieri* (L.): a highly valuable medic in al plant. *International Journal of Current Microbiology and Applied Sciences* **2**(12) 198-205.

Jain P, Khanna NK, Trehan N, Pendse VK and Godhwani JL (1994). Anti inflammatory effects of an Ayurvedic preparation, Brahmi Rasayana, in rodents. *Indian Journal of Experimental Biology* **32** 633-636.

Kaur J, Nautiyal K and Pant M (2013). In vitro propagation of Bacopa monnieri (L.) Wettst - A medicinally priced herb. International Journal of Current Microbiology and Applied Sciences 2(8) 131-138

Kapil SS and Sharma V (2014). In-Vitro Propagation of BacopaMonneri: An Important Medicinal Plant. *International Journal of Current Biotechnology* **2**(1) 7-10.

Kaur J, Nautiyal K and Pant M (2013). In vitro propagation of Bacopa monnieri (L.) Wettst A medicinally priced herb. International Journal of Current Microbiology and Applied Sciences 2(8) 131-138.

Kumari S, Starlin NM and Huxley AJ (2010). In vitro propagation of *Bacopa monnieri* (L.) - a wetland medic in al plant. *Journal of Basic and Applied Biology* **4**(3) 138-142.

Mehta J, Ansari R, Syedy M, Khan S, Sharma S, Gupta N, Rathore R and Vaishnav K (2012). An effective method for high frequency multiple shoots regeneration and callus induction of *Bacopa*

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2016 Vol. 5 (3) July-September, pp.17-23/Vijay et al.

Research Article

monnieri (L.) Pennell: an important medicinal plant. Asian Journal of Plant Science & Research 2(5) 620-626.

Mohanta YK and Sahoo S (2014). *In vitro* culture of highly valuable medicinal plant *Bacopa monnieri* (L.) Penn. for rapid and mass multiplication. *International Journal of Pharmaceutical Science Invention* **3**(1) 41-45.

Mohapatra HP and Rath SP (2005). *In vitro* studies of *Bacopa monnieri*-an important medicinal plant with reference to its biochemical variations. *Indian Journal of Experimental Biology* **43** 313-316.

Murashig T and Skoog F (1962). A revised medium for rapid growth and bioassays with tobacco tissue cultures. *Physiologia Plantarum* 15 473-497.

Nwokocha NJ, Umechuruba CI, Wokocha RC, Opara EU and Nwokocha JV (2015). Reduction of Seed-Borne Fungi of the Genus Aspergillus Associated with Egusi Melon Colocynthiscitrullus (L.) Seeds Using Chlorine Disinfectants – Implications on Seed Germination. *Journal of Agriculture and Sustainability* 7(1) 87-98.

Pandiyan P and Selvaraj T (2012). In vitro multiplication of *Bacopa monnieri* (L.)Pennell from shoot tip and nodal explants. *Journal of Agricultural Technology* **8**(3) 1099-1108.

Ramesh M, Saravanakumar RM and Pandian SK (2006). Benzyl amino purine and adenine sulphate induced multiple shoot and root induction from nodal explants of Brahmi, *Bacopa monnieri* (Linn.) Penn. *Green Page* 5(1) 44-51.

Rastogi S, Mehrotra BN and Kulshreshtha DK (1994). In: Deep publications (edition) *Proceedings of IV International Congress of Ethnobiology*, New Delhi, India 93.

Sairkar P, Chandravanshi MK, Shukla NP and Mehrotra NN (2009). Mass production of an economically important medicinal plant *Stevia rebaudiana* using *in vitro* propagation techniques. *Journal of Medicinal Plants Research* 3(4) 266-270.

Satyavati GV, Raina MK and Sharma M (1976). *Indian Medicinal Plants*, 1, (Indian Council of Medical Research, New Delhi, India).

Shankar G and Singh HK (2000). Anxiolytic profile of standardized brahmi extract. *Indian Journal of Pharmacology* **32** 152.

Sharma S, Kamal B, Rathi N, Chauhan S, Jadon V, Vats N, Gehlot A and Arya S (2010). *In vitro* rapid and mass multiplication of highly valuable medicinal plant *Bacopamonnieri*(L.)Wettst. *African Journal of Biotechnology* **9**(49) 8318-8322.

Showkat P, Zaidi Y, Asghar S and Jamaluddin S (2010). In vitro propagation and callus formation of *Bacopa monnieri* (L.) Penn. *Plant Tissue Culture and Biotechnology* **20**(2) 119-125.

Shrivastava N and Rajani M (1999). Multiple shoot regeneration and tissue culture studies on *Bacopa monnieri* (L.) Pennell. *Plant Cell Reports* **18** 919-923.

Srivastava S, Mishra N and Misra U (2009). Bacopamonniera -a Future Perspective. *International Journal of Pharmaceutical Sciences and Drug Research* **1**(3) 154-157.

Subashri B and Pillai YJK (2014). In vitro Regeneration of Bacopa Monnieri (L.) Pennel.-A Multipurpose Medicinal Plant. *International Journal of Pharmacy and Pharmaceutical Sciences* **6**(4) 559-563.

Tanveer A, Khan M and Shah F (2010). *In vitro* Micropropagation of *Brahmi-Bacopamonniera* (L.) Pennell A Step for Conservation. *Nanobiotechnica Universale* **1**(2) 139-150.

Tripathi YB, Chaurasia S, Tripathi E, Upadhaya A and Dubey GP (1996). Bacopa Monniera Linn as an antioxidant: mechanism of action. *Indian Journal of Experimental Biology* **34** 521-526.

Vohora SB, Khanna T, Athar M and Ahmad B (1997). Anagelesic activity of bacosine, a new tritepene isolated from Bacopamonniera. *Fitoterapia* **68** 361-365.

Williams R, Münch G, Gyengesi E and Bennett L (2014). Bacopa monnieri (L.) exerts antiinflammatory effects on cells of the innate immune system in vitro. Food & Function 5(3) 517-20.

Yusuf A, Rajesh KT, Nikhilesh S and Rao PS (2011). Effects of antioxidants and gelling agents on regeneration, in vitro conservation and genetic stability of *Bacopa monnieri* (L.) Pennell. *International Journal of Ayurvedic and Herbal Medicine* **1**(3) 51-67.