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CURE OF HEAVY METAL (CUSO₄) INDUCED ALTERATIONS IN AN EXPERIMENTAL MODEL, *BELLAMYA DISSIMILIS* BY CAFFEINE (1, 3, 7 - TRIMETHYLXANTHINE)

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ABSTRACT

The present study was conducted to evaluate the effectiveness of caffeine (1,3,7- Trimethylxanthine) in copper induced toxicity in an experimental model, the fresh water snail, *Bellamya dissimilis*. The effect on snails was studied under five groups. Group A was maintained as control, group B snails were exposed to chronic LC $_{50/10}$ dose of copper sulphate (0.041 ppm) for 21 days, while group C snails were exposed to respective chronic concentraction of copper sulphate along with 5 mg/lit of caffeine. Protein content in selected tissues of *Bellamya dissimilis*. from all groups were estimated after 7, 14 and 21 days. Snails from B groups were allowed to cure naturally while those of E were exposed to caffeine (5 mg/lit.). Protein contents in selected tissues from these D & E group snails were stuided after 7, 14 and 21 days. Significant decrease in protein content was observed in copper sulphate exposed snails as compared to control. The groups exposed to copper sulphate along with caffeine showed more protein content in the tissues than those exposed to heavy metal. Preexposed snails to copper sulphate showed to cure naturally. The probable role of caffeine is discussed in the paper.

Key Words: Caffeine, Protein Content, Copper, Bellamya Dissimilis.

INTRODUCTION

Heavy metals are recognized as a strong biotoxicants, becauseof their persistent nature and cumulative action to the aquaticflora and fauna (Sharma and Agrawal, 2005). The discharge of heavy metals by industries pose a serious water problem due to the toxic properties of these metals and their adverse effects on aquatic life. According to the survey conducted by Central Inland Fisheries Research Institute (CIFRI, 1981), these heavy metals are well kowan pollutants which are often encountered in many rivers of India, and there is every possibility of deterioration of water quality and hence including man and various organisms are presenting a potential threat for survival. Heavy metals are economic poisons used to control a wide range of animal and plant pests. The fresh water environment is becoming increasingly polluted throughout the biosphere with various heavy metals and as heavy metals are non-biodegradable, their concentration in the environment increases. The effect of heavy metals on animal life in fresh water is an important aspect of pollution and the information available on the physiological effects of exposure to different pollutants is meagre (Kleerkoper, 1974). These environmental pollutents bring about damage to different organs or disturb the physiological and biochemical processes within the organism.

In the aquatic invertebrate, Beaby and Eaves, (1983) observed that molluscscan accumulate higher concentration of metal ions than other groups of invertebrates. Among the heavy metals copper is important metal which is mostly used in the industries, paints and ceramics. The fertilizers are the main sources of copper, zinc and mercury which cause the pollution to the different media (Simkiss, 1984; Crop and Morgan, 1991). Intoxication of copper reduces growth, survival and rate of reproduction in the aquatic invertebrates.

Wood, (1974) classified metals into non-critical, very toxic and toxic metals. He classified arsenic, lead & mercury as very toxic heavy metals. These heavy metals enter into the body of animals including man through the non vegetarian and vegetarian diet, drinking water and air and accumulate in the tissues,

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usually react with proteins and interfer the physiological activities and thus increase the risk of life in various ways. They are difficult to remove from the body.

Micke Mc Laughlin, (2000) of CSIRO, Australia has found that coffee has capacity to bind with heavy metals. Heavy metal content of water was much reduced after addition of caffeine. Dissolved heavy metal ions are positively charged and coffee contains uncharged and negatively charged molecules, the metals ions might be taken out of solution by binding to negatively charged molecules in the coffee granules. Liguori, (1997) reported that,caffeine from coffee or other beverages is absorbed by the small intestine within 45 minutes of ingestion and then distributed throughout all tissues of the body. Metabolites of caffeine also contribute to caffeine's effects. Paraxanthine is responsible for an increase in the lipolysis process, which releases glycerol and fatty acids into the blood to be used as a source of fuel by the muscles. Theobromine is a vasodilator that increases the amount of oxygen and nutrient flow to the brain and muscles. Theophylline acts as a smooth muscle relaxant that chiefly affects bronchioles and acts as a chronotrope and inotrope that increases heart rate and force of contraction (Dews,1984). Maughan, (2003) sugested that, caffeine equivalent to 2–3 cups of coffee are administered to people who have not consumed caffeine during prior days, they produce a stimulation in urinary output

The molecules of coffee being small, it's chelate with heavy metal can be easily excreted out by the biological system. This property of caffeine indicates that caffeine can have the capacity to remove the heavy metals from the living organisms. However no attempt has been made to study the role of caffeine in heavy metal detoxification.

MATERIALS AND METHODS

The snails, *Bellamya dissimilis* were acclimatized to laboratory condition for 2-3 days and healthy active snails of approximately medium size and weight were chosen. These snails were divided into three groups, such as group A, B and C. The snails of group A were maintained as control. The snails from group B were exposed to chronic concentration (LC 50 value of 96 hr/10) of heavy metal salt, Copper sulphate (0.041 ppm) upto 21 days, while snails from group C were exposed to chronic concentraction of along copper sulphate with 5 mg/lit caffeine upto 21 days. After exposure for 21 days to heavy metals, the snails from group B were divided into two subgroups, such as D and E groups. The snails of group D were allowed for self cure naturally in normal water while the snails of group E were exposed to 5 mg/lit caffeine up to 21 days. During experimentation snails were fed on fresh water algae. The whole body, hepatopancreas and gonad of snails from all groups were collected after every seven days and were dried at 80^{0} C in an oven till constant weight was obtained. Protein contents from dried powders of different tissues of control and experimental animals were estimated by Lowry's method (Lowry *et al.*,1951).

RESULTS

Protein contents in different tissues of *Bellamya dissimilis* after exposure to Copper sulphate (0.041 ppm) along with caffeine and during recovery have been summarised in tables.

Table A shows that the protein contents in whole body, hepatopancreas and gonad *Bellamya dissimilis* in presence of copper sulphate (0.041 ppm) decreased with the increase in exposure period. The protein contents were more in heavy metal with caffeine exposed snails as compared to those exposed to only heavy metal salts for the corresponding period of exposure. Table B shows, that the snails preexposed to heavy metal salts showed fast recovery in the alteration of protein in presence of caffeine than those allowed to cure naturally. Therefore after studies the effect of caffeine on heavy metal it is proved that ,The caffeine posseses binding site to connect heavy metal salts and due to this effect of heavy metal is less in animals those exposed in caffeine as compared to those exposed only in heavy metal salts.

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Table A: Protein	Content in Selected Tissues of	Bellamya Dissimilis, after	r Chronic Exposure to
Heavy Metal Salt,	Copper Sulphate without and w	vith Caffeine.	

Treatment	Sr No.	Body Tissue [–]	The protein content (%) \pm S.D.		
			7 Days	14 Days	21 Days
(A) Control	i	W.B.	52.2 <u>+</u> 0.0033	51.5 <u>+</u> 0.0034	50.2 <u>+</u> 0.0091
	ii	H.	36.0 <u>+</u> 0.0044	<u>34.2+</u> 0.0074	33.7 <u>+</u> 0.0088
	iii	G.	32.5 <u>+</u> 0.0052	31.7 <u>+</u> 0.0079	30.0 <u>+</u> 0.0089
(B) 0.041 ppm CuSO ₄	i	W.B.	$50.7 \pm 0.0038^{***}$	$49.0 \pm 0.0008^{***}$	$47.2 \pm 0.0057^{***}$
			-2.958°	-5.102 •	-6.355°
	ii	H.	$34.5 \pm 0.0034^{***}$	32.2 <u>+</u> 0.0047 ^{***}	31.7 <u>+</u> 0.0160 ^{***}
			- 4.347°	- 6.211°	- 6.309 •
	iii	G.	30.7 <u>+</u> 0.0091 ^{***}	$29.5 \pm 0.022^{***}$	$28.2 \pm 0.0368^{***}$
			- 5.863 °	- 7.457 •	- 6.382 •
(C) 0.041ppm CuSO ₄ + 5mg/lit Caffeine	i	W.B.	51.7 <u>+</u> 0.0047 ^{***}	$50.5 \pm 0.0101^{***}$	49.7 <u>+</u> 0.0091 ^{***}
			-0.967•, +1.934 ^Δ	$-1.980^{\bullet}, +2.970^{\Delta}$	$-1.006^{\bullet}, +5.030^{\Delta}$
	ii	H.	$35.2 \pm 0.0017^{***}$	33.5 <u>+</u> 0.135 ^{***}	33.2 <u>+</u> 0.0097 ^{***}
			$-2.272^{\bullet}, +1.988^{\Delta}$	$-2.0.89^{\bullet}, +3.880^{\Delta}$	$-1.506^{\bullet}, +4.518^{\Delta}$
	iii	G.	31.7 <u>+</u> 0.0079 ^{***}	$30.7 \pm 0.0065^{***}$	30.0 ± 0.0089^{NS}
			-2.523 [•] , +3.154 [△]	-3.257°, +3.908 [∆]	$-0.000^{\bullet}, +6.000^{\Delta}$

Table B :after 21 Days Exposure to 0.041 ppm Cuso4

Treatmont	Sr	Body	The protein content (%) \pm S.D.		
Treatment	No.	Tissue	28 Days	35 Days	45 Days
(D) Normal Water	i	W.B.	$48.5 \pm 0.0072^{***}$	$49.2 \pm 0.0074^{***}$	50.0 ± 0.0089^{NS}
			$-7.628^{\bullet}, +2.680^{\Box}$	$-4.674^{\bullet}, +4.065^{\Box}$	- 0.400^{\bullet} , + 5.600^{\Box}
	ii	H.	$32.5 \pm 0.0052^{***}$	$33.5 \pm 0.0071^{***}$	34.2 ± 0.033^{NS}
			- 10.760^{\bullet} , + 2.461^{\Box}	$-2.089^{\bullet}, +5.373^{\Box}$	$+1.461^{\bullet}, +7.309^{\Box}$
	iii	G.	$29.7 \pm 0.0079^{***}$	$30.0 \pm 0.0089^{***}$	31.5 ± 0.004^{NS}
			$-9.427^{\bullet}, +5.050^{\Box}$	$-5.666^{\bullet}, +0.000^{\Box}$	$+4.761^{\bullet}, +10.476^{\Box}$
(E) Normal Water + 5mg/ lit. Caffeine	i	W.B.	$49.7 \pm 0.0091^{***}$	$50.5 \pm 0.101^{***}$	51.5 ± 0.0034^{NS}
			$-5.030^{\bullet}, +5.030^{\Box}$	- 1.980^{\bullet} , + 6.534^{\Box}	$+2.524^{\bullet}, +8.349^{\Box}$
	ii	H.	33.2 <u>+</u> 0.0097 ^{***}	34.7 <u>+</u> 0.0079 ^{***}	35.7 ± 0.0089^{NS}
			- 8.433°, + 4.518 $^{\Box}$	$+ 1.440^{\bullet}, + 8.645^{\Box}$	$+5.602^{\bullet}, +11.204^{\Box}$
	iii	G.	30.2 <u>+</u> 0.0079 ^{***}	31.5 <u>+</u> 0.0048 ^{***}	32.0 ± 0.0169^{NS}
			- 7.615°, + 6.622 $^{\Box}$	$-0.634^{\bullet}, +10.476^{\Box}$	+ 6.832°, + [□]
W.B Whole Body		N.S Non Significant		• - Compared with respective A	
H Hepatopancreas		* - P < 0.005		Δ - Compared with respective B	
G - Gonads		**	P - $P < 0.01$ \square - Compared with respective		l with respective 21
		***	* - P < 0.001	days of B	

DISCUSSION

The change in biochemical composition of an organ due to heavy metal stress indicates the change in activity of an organism. It reflects light on the utilisation of their biochamical energy to counteract the

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toxic stress. Heavy metal salts affect the metabiolism of the fresh water snails, *Bellamya dissimilis*. Alterations in metabolic pocesses, following exposure to heavy metal stress have been always used as an indicator of stress. But there is a vast difference in the pattern & metal induced physiological alterations from metal to metal & animal to animal.Protein content in the tissue of animal is an important essential organic constituent which plays a vital role in the cellular metabolism. All enzymes are proteins in nature and they control subcellular functions and accelarate the rate of metablic action in the body of organism. Ramanarao and Ramamurthi, (1978) studied the protein content in the tissue of *Pila globosa* after exposure to pesticide.

In present study, in the *Bellamya dissimilis* the protein contents in the selected tissues was decreased in chronic concentration of copper sulphate as compared to the control and LC50/10 concentration with 5 mg/lit. According to Abel, (1974) the decrease of protein may be due to alterations of membrane permeability. The depletion in the protein content was reported from the muscles of fish, *Clarias batrachus* after treatment with pesticide by the Yagana Bano et al., (1981). Nagabhushanm and Kulkarni, (1979) studied variation in protein metablism in *Barytelephusa cunicularis*. Joseph et al., (1987) observed the effect of copper on biochemical composition of *Cyprinus carpio* and found that total protein content of the brain, liver and muscles was declined.

Mukherjee and Sinha, (1993) studied the effect of heavy metal toxicity on haematological and biochemical aspect in the fresh water major carps, *Labeo rohita*. Katticaran et al., (1995) studied the copper induced alterations in total carbohydrate and protein level in the bivalve, *Sunetta scripta*.

In present stress, ionic copper sulphate might have caused severe disturbances of the metabolism in the animal. Chronic exposure of copper sulphate alone showed a remarkable decrease in protein content in *Bellamya dissimilis* than the chronic doses of heavy metal salts with caffeine.

Starvic, (1994) after the study of role of chemopreventer in human diet suggested that most of the chemopreventive strategies have been based on the modification of metabolism at one or many steps such that these agents can block the metabolic activition or enhance the detoxification . Caffeine is wellknown nervous system stimulant but besides it, it is now observed that it has antioxident activity. This activity of caffeine can protect the damage of tissues chemicals & genetic materials from heavy metal generated free oxygen radicals.Oral administration of tea has been found to moderately inhance the activities of lipid Peroxidase .catalase, glutathione-s-transferase which in turn protect against cancer by blocking the reaction of electrophilic carcinogens with cellular micromolecules (Madal and Maity, 1999).Caffeine being water soluble and common cheaper beverage, it will be cheapest preventive and curative medicine. Takayamas, (1982) long term study on the effect of caffeine in wister rats, has proved that caffeine belongs to a group of compound known as methylxanthine and it is non carcinogenic in animal model. It has also been reported to antagonize the carcinogenic effects of chemicals in vitro. Under in vitro condition caffeine has been reported to enhance or inhibit tumorigenesis induced by various carcinogenic agents, mercury, arsenic and lead are the known carcinogenics.Wattenberg (1992) reported that any compound that can block the metabolic activation step, scavenge the reactive inter mediate or enhance detoxification would be potential chemopreventive agent. Chung Fung-Lung (1999) reported that caffeine when given in drinking water at a concentration identical to that found in 2% tea was able to inhibit lung tumours induced by 4-(Methylnitrosoamino)-1-(3-pyridyl)-1 furane (NNK). Hosakas et al.,(2001) has observed that caffeine inhibite hepatocarcinogenesis induced by 2- acetylaminoflurene.

Gandhi and Khanduja, (1992) studied the action of caffeine in altering the carcinogen activiting and detoxofying enzymes in mice and reported an induction of xenobiotic detoxifying enzyme as an additional mechanism by which plant product may act as anticarcinogens, since this induction of detoxifying enzyms is capable of competing with steps in xenobiotic activation. Caffeine have been found to increase glutathione synthetase and reduced glutathione in liver and lungs of mouse (Gandhi and Khanduja, 1992). Lu *et al.*, (2001) studied the stimulatory effect of oral administration of green tea or Caffeine on ultraviolet light induced alterations and suggested that green tea & caffeine inhibits UV-induced carcinogenesis.Matsumura *et al.*, (2000) reported that, the ATP – requring priming stage is

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lacking in the process of caffeine induced exocytosis in bovine adrenal chromaffin cells. Hove-madsen, (1999) suggested caffeine exposure incressed pulse duration to 85m/s and slowed the inactivation of the Ca2+ current (Ica). Leoty *et al.*, (2001) found that caffeine stimulates the reserve mode of NA(Sup+)/Ca(SUP2+) exchanger in ferrete ventrecular muscles. and indicated that the increase in resting tension following exposure to caffeine was mediated by Na+/Ca2+ exchanger, Which represents an additional element of coplexity in caffiene action on cardiac muscles. Massey *et al.*, (1993) indicated the increased urinary excretion of calcium, magnesium, sodium and chloride after oral doses of caffeine which indicates the chelated caffeine with heavy metal is excretable. In September (2001), Women's Health Weekly also reported that, the caffeine in the drinks was primarily responsible for excess calcium excretion.

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REFERENCES

Abel PD (1974). Toxicity of synthetic detergents to fish and aquatic vertebrates. *Journal of Fish Biology* 6 279–298.

Beaby A and Eaves SL (1983). Short term changes in (Pb, Zn and Cd concentrations of the garden Snail Helixaspersa (Muller) from a central London Car Park. *Environmental pollution (series A)* **30** 233-244.

Chung Fung- Lung (1999). The prevention of lung cancer induced by a tabacco specific carcinogen in odentsby green & black tea. *Proceedings* of the *Society* for *Experimental Biology* and *Medicine journal* 220 - 224.

Dews PB (1984). Caffeine: Perspectives from Recent Research. Berlin: Springer-Valerag.

Gandhi RK and Khanduja KL (1992). Action of caffeine in altering the carcinogen activating and detoxifying enzymes in mice. *Journal of Clinical Biochemistry* 25 12 - 19.

Hosaka SS, Kawa S, Akoi Y, Tanaka E, Yoshizawa K, Karasawa Y, Hosaka N, and Kiyosawa K (2001). Hepato carcinogenesis inhibition by caffeine in ACI rats treated with - acetylamino flurene. *Food and chemical Toxicology journal* (39) 557.

Hove-Madsen, Leif, Anna Llach and Liuis Tort (1999). Quantification of calcium release from the sacoplasmic reticulum in rainbow trout atrial myocytes, Pffuergers Archi. *European Journal of Physiology* **438**(4) 545-552.

Joseph et al. (1987). Chronic toxicity of copper on the biochemical composition of some tissues of the scale carp, Cyprinus carpiocommunis. Proceedings of the National Conference on .Environment Impact on Biosystem 263–267.

Katticaran CM, Mohammed Saih KY and Joseph PS (1995). Copper induced alterations in total carbohydrates and proteion levels in bivalve, *Sunetta scripta* (bivalvia). *Indian Journal of Marine*

Sciences **24**(3) 171–174. **Liguori A, Hughes JR, Grass JA (1997).** Absorption and subjective effects of caffeine from coffee, cola and capsules. *Pharmacology Biochemistry and Behaviour* **58**(3) 721–6.

Leoty C, Huchet- Cadiou C, Talon S, Choisy S and Hleihel W (2001). Caffeine stimulates the reserve mode of NA (SVP+) /CA (SUP 2+) exchanger in ferret ventricular muscles. *Acta physiological scandinavica* 172(1).

Lu Yau – Ping, You-Rong Lou, Xiang Hong Li and Jian Guoxic (2000). Stimulatory effect of oral administration of green tea or caffeine on Uv- light-induced increases in epidermal wildtype p53 and p21. *Cancer Reserch* **60**(17) 4785-4791.

Maughan RJ, Griffin J (2003). Caffeine ingestion and fluid balance: a review. *Journal of Human Nutrition and Diet* 16(6) 411–420.

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Matsumura C, H-Kuwashima and T. Kimura (2000). Lack of Ca2+ & ATP – dependent priming stage in caffeine induced exocytosis in bovine adrenal Chromaffin cells. *Journal of Autonomic pharmacology* 20(1) 31-36.

Massey, Linda K, Whiting and Susan J (1993). Caffeine, urinary calcium, calcium metabolism and bone. *Journal of Nutritions* 123 1611-1649.

Mukharjee JK and Sinha GM (1993). Cadmium toxicity on haematological and biochemical aspects in an Indian freshwater major carps, *Labeo rohita* (Hamilton). *Journal of Freshwater Biology* 5(3) 245–251. Nagabushanam R and Kulkarni GK (1979). Effect of thermal acclimation on protein metabolism in

Nagabushanam R and Kulkarni GK (1979). Effect of thermal acclimation on protein metabolism in the fresh water crab, *Barytelphusa cunicularis*. *Bioreserch* **3**(1) 9-13.

Sharma RJ and Agrawal M (2005). An overview of Biological effects of heavy metals. *Journal of Environmental Biology* 26 301-338.

Simkiss K (1984). Invertebrates give neutralization metals toxics. Mundo Cientifica 39 864-866.

Starvic CB (1994). Role of Chemopreventeres in human diet. Clinical Biochemistry 27 319-325.

Ratankar Dhar, Bajan Kr and Biswas et al. (1997). Groundwater arsenic calamity in Bangladesh 48-59.

Ramanarao K and Ramamurthi R (1978). Studies on metabolism of the apple snail, *Pila*

globosa.(Swainson)in relation to pesticide impact. *Indian Journal of Heredity* 10-11. **Takayama S** (1982). long term study on the effect of caffeine in wister rats, Gann (73) 365.

Wattenberg LW *et al* (1992). Chemoprevention of cancer by naturally occuring and synthetic

Compounds. Ann Arbor MI: CRC press 19.

Yagana Bano, Seikh, mjad Ali and Taric Hameed (1981). Effect of sublethal concentration of DDT on muscle constituents of an air breathing catfish, *Clarius batrachus*. *Proceedings of IndianAcademy of Science* (animal science) 90(1) 33-39.