# ELECTROPHORETIC ANALYSIS OF EXCRETORY/SECRETORY ANTIGENS OF *GASTROTHYLAX CRUMENIFER* FROM BUFFALO

## Arunkumar S.<sup>1</sup>, Prakash Krupakaran R.<sup>2</sup>, Balamurugan T.C.<sup>2</sup> and Guru D.V. Pandiyan<sup>2</sup>

<sup>1</sup>Department of Veterinary Parasitology, Veterinary College and Research Institute, Orathanadu-614 625, Thanjavur (Dist), Tamil Nadu <sup>2</sup>Department of Veterinary Physiology and Biochemistry, Veterinary College and Research Institute, Orathanadu-614 625, Thanjavur (Dist), Tamil Nadu \*Author for Correspondence

#### ABSTRACT

Live adult *Gastrothylax crumenifer* worms were collected from buffaloes slaughtered at local abattoirs and washed several times with PBS (pH 7.4). Live intact adult flukes were suspended in DPBS (pH 7.2) and incubated at 37°C in a BOD incubator for 8 hrs. The fluid was centrifuged at 7000 rpm for 30 min at 4°C and the supernatant was designated Excretory and Secretory (E/S) antigens. On Sodium dodecyl sulphate-polyacrylamide gel electrophoresis (SDS-PAGE) analysis, seven major bands *viz.*, 240, 220, 180,130, 92, 72 and 30 kDa and three minor bands having molecular weight (MW) lesser than 30 kDa size were observed.

Keywords: Gastrothylax Crumenifer, E/S Antigens, SDS-PAGE, Buffalo

## INTRODUCTION

Paramphistomosis of livestock is considered as a disease of great economic importance. It is widely prevalent in domestic ruminants in India and several other countries, resulting in heavy losses in terms of morbidity and reduced wool, meat, and milk production (Manna, 1994 and Hassan *et al.*, 2005). A number of investigators have been trying to develop specific serodiagnostic tests for early detection of a related trematode infection in animals (Yadav *et al.*, 2005; Raina *et al.*, 2006). Some of the methods have been tested against experimental and field infections of cattle and buffaloes, but, little effort has been made in immunodiagnosis of ruminal amphistomes. Hence, the study was carried out to analyze the protein profiles of in excretory/ secretory (E/S) antigens of *Gastrothylax crumenifer* through SDS-PAGE.

## MATERIALS AND METHODS

Live, mature *Gastrothylax crumenifer* flukes were collected from the rumen of Indian water buffaloes (*Bubalus bubalis*), slaughtered at the local abattoirs of Pattukkottai and Thanjavur. The flukes were thoroughly washed in Phosphate Buffer Saline without glucose, pH 7.4 and pre maintained at 37°C. After careful preservation in PBS, the worms were immediately transferred to the laboratory for further processing. The flukes were identified based on specific morphological characters (Soulsby, 1981). Excretory-secretory antigens (ES-Ag) were prepared as per the procedure described by Saifullah *et al.*, (2011) with minor modifications. Live intact adult flukes were weighed and suspended in DPBS (pH 7.2) and were incubated at 37°C in a BOD incubator for 8 hr. Then, the fluid was centrifuged at 7000 rpm for 30 minutes at 4°C and the supernatant collected was designated as E/S antigens. The E/S antigens was further lyophilized in a centrifugal freeze-dryer and then it was reconstituted in DPBS and stored at -20°C till further use. The total protein content of the samples was estimated (Lowry *et al.*, 1951). SDS-PAGE analysis of E/S antigens was carried out as per the method described by Laemmli (1970) and the gels were silver stained by the method of Merril *et al.*, 1981.

#### **RESULTS AND DISCUSSION**

In the present study, the total protein concentration was 0.163 mg/mL. Each gel well was loaded with 80  $\mu$ L of E/S sample. 10 % SDS-PAGE (discontinuous method) under non reducing conditions was carried out at 100V for 8 hours. Then, the gel was silver stained by adopting the method of Laemmli (1970). The

Cibtech Journal of Bio-Protocols ISSN: 2319–3840 (Online) An Open Access, Online International Journal Available at http://www.cibtech.org/cjbp.htm 2014 Vol. 3 (3) September-December, pp.24-26/Arunkumar et al. **Research Article** 

electrophoretogram was studied using the protein marker (low molecular weight Genei, Bangalore) 3.5 to 205 kDa. Seven prominent bands ranging from 240 kDa to 5 kDa were observed as shown in the figure.



Figure 1: SDS-PAGE analysis of E/S antigens of Gatrothylax crumnifer (Silver staining) Lane 1- E/S antigens of Gatrothylax crumnifer Lane 2- Protein markers (Low molecular weight)

On SDS-PAGE analysis, seven major bands *viz.*, 240, 220, 180,130, 92, 72 and 30 kDa and three minor bands having molecular weight (MW) lesser than 30 kDa were observed. Fainter bands around 160 kDa were also observed. Among the prominent bands, the intensity was stronger for higher MW bands (240, 220 and 180 kDa). The intensities of low MW prominent bands were weaker and the separation was not very discrete suggesting the presence of non proteinaceous complex nature of these proteins. Saifullah *et al.*, (2000) reported the presence of heterogeneous population of varying MW ranging from 14 to 205 kDa in eight partially purified fractions of somatic extracts of *Gastrothylax crumenifer*.

Saifullah *et al.*, (2011) reported SDS-PAGE profile of purified fractions of *Gastrothylax crumenifer* containing 8-12 polypeptides having MW. less than 14 to 165 kDa and reported the presence of only three major bands at 105, 141 and 165 kDa. In our study also, we observed bands having MW in the range of < 5kDa. Four prominent bands having MW > 130 kDa and the presence of several polypeptide bands in the range of < 29 to > 205 kDa, which corroborated with the results of Ahmed *et al.*, (2004). However, the slight variations in the relative MW of the polypeptides may be due to the influence of season on the reproductive cycle of parasite as reported earlier by Hanna *et al.*, (1988) and the geographical location of the parasite. Hence, further studies on purification and characterization of E/S antigens of *G. crumnifer*, which could help to develop specific serodiagnostic test for earlier detection of Paramphistomosis in buffaloes.

#### REFERENCES

Ahmad G, Saifullah MK and Nizami WA (2004). Partial purification and characterization of *Gigantocotyle explanatum* somatic antigens. *Journal of Helminthology* **78**(2) 95-9.

Hanna REB, Willamson DS, Mattison RG and Nizami WA (1988). Seasonal reproduction in *Paramphistomum* and *Gastrothylax crumenifer* rumen amphistomes of the Indian water buffalo and comparison with the biliary paramphistome *Gigantocotyle explanatum*. *International Journal for Parasitology* **18** 513–521.

Hassan SS, Kaur K, Joshi K and Juyal PD (2005). Epidemiology of paramphistomosis in domestic ruminants in different district of Punjab and other adjoining areas. *Journal of Veterinary Parasitology* **19** 43-546.

Laemmli UK (1971). Cleavage of structural proteins during the assembly of the head of bacteriophage T4. *Nature* 227 680-685.

Cibtech Journal of Bio-Protocols ISSN: 2319–3840 (Online) An Open Access, Online International Journal Available at http://www.cibtech.org/cjbp.htm 2014 Vol. 3 (3) September-December, pp.24-26/Arunkumar et al. **Research Article** 

Lowry OH, Rosebrough NJ, Farr AL and Randall RJ (1951). Protein measurement with the folin phenol reagent. *The Journal of Biological Chemistry* 193(1) 265-275.

Manna AK, Pramanik S and Mukherjee GS (1994). Incidence of paramphistomiasis in west Bengal. Indian Journal Of Animal Health 33 87-89.

Merril CR, Goldman D, Sedman SA and Ebert MH (1981). Ultrasensitive stain for proteins in polyacrylamide gels shows regional variation in cerebrospinal fluid proteins. *Science* 211 1437-1438.

**Raina OK, Yadav SC, Sriveny D and Gupta SC (2006).** Immunodiagnosis of bubaline fasciolosis with *Fasciola gigantica cathepsin-L* and recombinant cathepsin L 1-D protease. *Acta Tropica* **98** 145–151.

Saifullah MK, Ahmad G and Abidi SM (2011). Isolation and partial characterization of excretory/ secretory antigens of *Gastrothylax crumenifer*. *Veterinary Parasitology* **180**(3-4) 232-6.s

Saifullah MK, Ahmad G, Nizami WA and Abidi SM (2000). Partial purification and characterization of *Gastrothylax crumenifer* somatic *antigens*. *Veterinary Parasitology* **89**(1-2) 23-9.

Souls by EJL (1981). *Helminths, Arthropods and Protozoa of Domesticated Animals*, Seventh edition (Bailliere and Tindal, London).

Yadav SC and Gupta SC (1995). Immunodiagnostic moieties in somatic and excretory/ secretory antigens of *Fasciola gigantica*. *Indian Journal of Experimental Biology* 33 824-828.