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PHENOTYPIC DETECTION OF METALLO BETA LACTAMASE IN GRAM NEGATIVE BACTERIAL ISOLATES

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ABSTRACT

Carbapenem Resistance due to the production of metallo-beta-lactamases (MBL) in Gram-negative organisms is an increasing international public health problem. Emergence of MBL producing organisms is alarming and reflects the excessive use of carbapenems. The problem of MBL producing strains was originally confined to *Pseudomonas* and *Acinetobacter*. However, carbapenem resistance has been observed in members of Enterobacteriaceae family due to spread of MBL genes. The present study was aimed at determining the prevalence of metallo-beta-lactamases (MBL) production in gram negative bacterial isolates obtained from various clinical isolates. A total of 1356 non-repeat isolates of gram negative bacterial isolates obtained from various clinical samples were processed by conventional methods. Out of these 1356 isolates, only those which were resistant to meropenem by the E-test method were included in this study. Among these resistant strains, MBL production was determined by the combined disk test (CDT). Out of 1356 non-repeat isolates of gram negative bacterial isolates, 112 isolates showed resistance to meropenem by the E-test method. Out of which 74.11% of the isolates turned out to be MBL producers by the CDT method. In this study, 85% of *Pseudomonas* species, 75.44% of *Acinetobacter*, 66.67% of *Klebsiella*, 50% each of *E.coli* and *Citrobacter* species were MBL positive. The maximum number of MBL producers were reported from respiratory samples followed by plastic devices. The higher prevalence rate of MBL producing organisms in the present study focuses on the need for strict implementation of infection control practices and antibiotic restriction policies to avoid excessive use of carbapenems and other broad-spectrum antibiotics.

Keywords: Carbapenems, Metallo-beta-lactamases (MBL), Gram negative bacteria, Meropenem, Combined disk test (CDT).

INTRODUCTION

The introduction of carbapenems into clinical practice was of great help in the treatment of serious bacterial infections caused by β -lactam resistant bacteria (Varaiya *et al.*, 2008). Carbapenems are often used as antibiotics of last resort for treating infections due to multidrug-resistant gram-negative bacilli, because they are stable even in response to extended-spectrum and AmpC β -lactamases (Lee *et al.*, 2003). The emergence of acquired metallo- β -lactamases (MBLs) in Gram-negative bacilli is becoming a therapeutic challenge, as these enzymes usually possess a broad hydrolysis profile that includes all β -lactam antibiotics with the exception of monobactams (Galani *et al.*, 2008). Unlike carbapenem resistance due to several other mechanisms, the resistance due to MBL and other carbapenemase production has a potential for rapid dissemination, as it is often plasmid mediated. (Walsh *et al.*, 2005) In recent years, MBL genes have spread from *P. aeruginosato* members of the Enterobacteriaceae family (Peleg *et al.*, 2005).

In addition to their resistance to all β -lactams, the MBL producing strains are frequently resistant to aminoglycosides and fluoroquinolones. MBL producing isolates are also associated with a higher morbidity and mortality (Walsh *et al.*, 2005). Moreover the treatment alternatives are unavailable or expensive/ toxic with poor outcome (Marra *et al.*, 2006).

Based on molecular studies, carbapenemhydrolyzing enzymes are classified into four groups A, B, C and D. The MBLs belong to group B and are enzymes requiring divalent cations as cofactors for enzyme activity, being inhibited by the action of a metal ion chelator (Ambler *et al.*, 1980). Although PCR-based

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genotyping remains as the golden standard for MBL detection and classification, its use is mainly restricted to research purposes (Khosravi *et al.*, 2012).

Several non-molecular techniques have been studied, all taking advantage of the enzyme's zinc dependence by using chelating agents, such as EDTA or 2 mercaptopropionic acid, to inhibit its activity. (Franklin *et al.*, 2006). There is a global increase in the prevalence of MBL-producing non-fermenting bacilli and Enterobacteriaceae (Walsh *et al.*, 2005; Galani *et al.*, 2008). Therefore this study was done to find the prevalence of Metallo beta lactamases among the gram negative bacteria isolated from various clinical samples.

MATERIALS AND METHODS

It was a prospective study conducted between October 2012-September 2013 at the department of microbiology of Dr B.R Ambedkar Medical College, Bangalore. A total of 1356 non-repeat isolates of gram negative bacteria were included in this study. The isolates were identified by conventional methods (Collee, 1999). Repeated isolates from the same patient were excluded. Antibiotic susceptibility testing was performed by the disc diffusion method using Mueller–Hinton agar (HIMEDIA) as per the CLSI 2012 guidelines. The MIC of meropenem was determined by the E test, according to the manufacturer's recommendations (BioMérieux, Marcy l'Etoile, France) and interpreted according to the CLSI 2012. Only meropenem resistant isolates were included in this study. These isolates were also resistant to other carbapenems and third generation cephalosporins. *E. coli* ATCC 25922 and *P. aeruginosa* ATCC 27853 were used as controls.

The meropenem resistant isolates were then subjected to the combined disk test (CDT) for the detection of MBL.

Combined disktest (CDT):

The MEROPENEM-EDTA combined disk test was performed as described by Yong *et al.*, 2002. Test organisms were inoculated on to plates with Mueller Hinton agar as recommended by the CLSI. Two 10 µg meropenem disks (Becton Dickinson) were placed on the plate, and appropriate amounts of 10 µL of EDTA solution were added to one of them to obtain the desired concentration (750 µg). The inhibition zones of the meropenem and meropenem-EDTA disks were compared after 16 to 18 hours of incubation in air at 35°C. In the combined disc test, if the increase in inhibition zone with the meropenem and EDTA disc was ≥ 7 mm than the meropenem disc alone, it was considered as MBL positive.

RESULTS

Out of which 1356 non-repeat isolates of gram negative bacterial isolates, 112 isolates showed resistance to meropenem by the E-test method. 57 out of 112 isolates screened were *Acinetobacter* species, 40 were *Pseudomonas* species, 9 were *Klebsiella*, 4 were *E.coli* and 2 were *Citrobacter* species.

Among the resistant strains, 85% of *Pseudomonas* species, 75.44% of *Acinetobacter*, 66.67% of *Klebsiella*, 50% each of *E.coli* and *Citrobacter* species were MBL positive by the Combined disk diffusion test (Table 1).

Table 1: Distribution of MBL producing organisms

Organism isolated	Meropenem resistant(%)	MBL positive by CDT (%)
<i>Pseudomonas</i>	40 (35.71%)	34 (85%)
<i>Acinetobacter</i>	57 (50.9%)	43 (75.44%)
<i>Klebsiella</i>	9 (8.06%)	6 (66.67%)
<i>E.coli</i>	4 (3.57%)	2 (50%)
<i>Citrobacter</i>	2 (1.8%)	1 (50%)
	112	86 (74.11%)

The maximum number of MBL positive organisms were isolated from the respiratory samples which included sputum and endotracheal aspirates followed by plastic devices which included central and peripheral venous catheters and urinary catheters (Table 2 and Chart 1).

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Table 2: Distribution of MBL producing bacterial isolates in various clinical samples :

	Respiratory samples	Pus	Urine	Blood	Genital samples	Plastic devices	Total
<i>Acinetobacter</i>	17	7	2	3	2	12	43
<i>Pseudomonas</i>	12	5	2	2	1	12	34
<i>Klebsiella</i>	4	1	1	0	0	0	6
<i>E.coli</i>	0	0	0	1	0	1	2
<i>Citrobacter</i>	0	1	0	0	0	0	1
Total	33	14	5	6	3	25	86

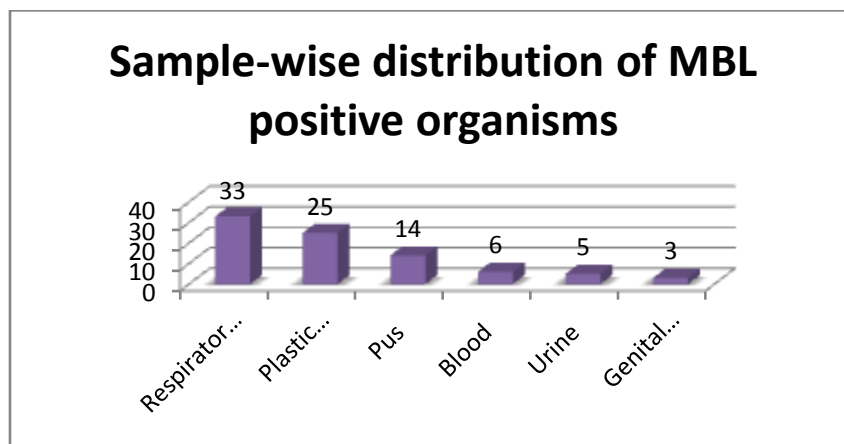


Chart 1: Sample-wise distribution of MBL positive organisms

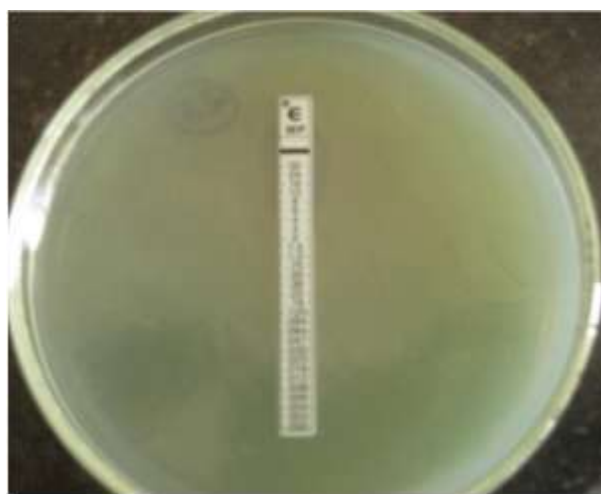


Figure 1: Meropenem E-test: *Pseudomonas aeruginosa* showing MIC ≥ 32 $\mu\text{g/ml}$

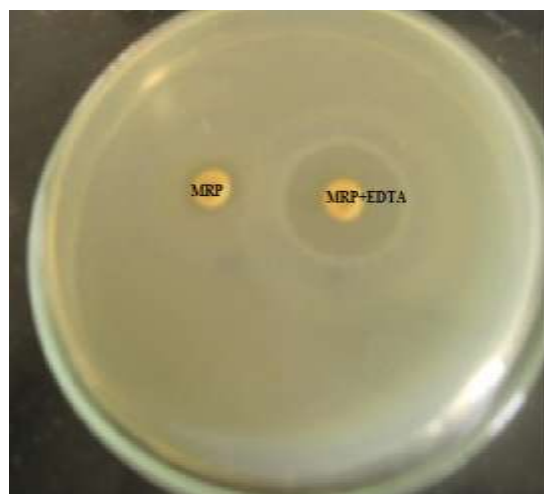


Figure 2 : Combined disk test (CDT) : Zone of inhibition of MRP+EDTA disc is ≥ 7 mm than that of MRP disc alone

DISCUSSION

Carbapenem Resistance due to the production of metallo-beta-lactamases (MBL) in Gram-negative organisms is an increasing international public health problem (Walsh *et al.*, 2005; Cornaglia *et al.*, 2007). Genes encoding for MBL were shown to be carried on large transferable plasmids or were associated with transposons, allowing horizontal transfer of these MBL genes among different bacterial genera and species (Pitout *et al.*, 2007).

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Carbapenemhydrolyzing enzymes are most commonly seen in nonfermenter gram negative organisms (non enterobacteriaceae) i.e. *Pseudomonas* and *Acinetobacter*. However, in the recent years there is an increasing incidence of these enzymes in Enterobacteriaceae family as well (Balan *et al.*, 2012).

In our study, *Acinetobacter* species showed maximum resistance to meropenem (50.9%). Similar resistance rates have been reported by other studies (Manikal *et al.*, 2000; Noyal *et al.*, 2009). Even higher rates have been reported by Nazmul *et al.*, 2012 (92.5%). *Pseudomonas* species showed resistance of 35.71% which also correlates with studies done by Bhat *et al.*, 2013 followed by *Klebsiella* species which showed a resistance rate of 8.06%. This is in concordance with Hodiwala *et al.*, 2013 and Pandya *et al.*, 2011.

The maximum number of these resistant isolates was obtained from respiratory samples which included endotracheal aspirates and sputum followed by plastic devices (catheter tip), pus, blood and urine indicating that the use of indwelling medical devices plays an important role in the spread of infective agents. Other studies have also reported a higher rate of resistant isolates from respiratory samples (Jesudasan *et al.*, 2005; Kaleem *et al.*, 2010).

Few studies have reported that 'Imipenem-EDTA combined disk test' as the most sensitive method for detection of MBL production in gram negative bacilli (Franklin *et al.*, 2006 and Pandya *et al.*, 2011). Hence we followed the combined disk method using meropenem- EDTA for the detection of MBL in the gram negative organisms. 74.11% of the meropenem resistant gram negative bacterial isolates were MBL positive in our study. Kaleem *et al.*, 2010 in Pakistan reported 78 % of the gram negative bacterial isolates as MBL positive.

Pseudomonas species showed the highest MBL production rate accounting to 85%, followed by *Acinetobacter* species (75.44%) and *Klebsiella* (66.67%). Pandya *et al.*, 2011 also observed a similar pattern of MBL resistance rate in their study on Gram negative bacterial isolates.

83.8% of *P. aeruginosa* were MBL positive in study done by Gupta *et al.*, 2013 and Hemalatha *et al.*, 2005 also reported 87.5 % MBL rate in *Pseudomonas*.

Acinetobacter species exhibited MBL production rate of 75.44% which correlates with study done by Tellis *et al.*, 2013. Higher rates (84% and 96%) have been observed by other studies in Pakistan (Kaleem *et al.*, 2010; Irfan *et al.*, 2008 respectively). Whereas studies done by Noyal *et al.*, 2009 and Anil *et al.*, 2011 in India have reported lower prevalence of MBL in *Acinetobacter* species (6.5% and 21% respectively). 66.67% of the *Klebsiella* isolates were MBL positive. These results were comparable to a study done by Jain *et al.*, 2012.

To conclude, the high rate of MBL producing gram negative bacteria in this study emphasizes on the need for active surveillance in the microbiology labs for the detection of these resistant strains and also stresses on the judicious use of carbapenems to prevent the spread of resistance. Different phenotypic methods for detection of these carbapenemases are available, but controversies exist regarding the choice of optimal laboratory method. Microbiology laboratories must be prepared to screen for MBL-producing isolates by a low cost, convenient and sensitive procedure.

REFERENCES

- Ambler RP (1980).** The structure of beta-lactamases. *Philosophical transactions of the Royal Society of London, Series B, Biological sciences* **289** 321- 31
- Anil VK, Vishnu SP, Kavitha R, Dinesh and Shamsul. (2011).** The phenotypic detection of carbapenemase in the meropenem resistant *Acinetobacter calcoaceticus-baumannii* complex in a tertiary care hospital in south India. *Journal of Clinical and Diagnostic Research* **5** 223-26.
- Balan K, Sireesha P and CR Setty (2012).** Study to detect incidence of carbapenemase among Gram negative clinical isolates from tertiary care hospital. *International Organization of Scientific Research Journal of Dental and Medical Sciences (JDMS)* **1** (6) 08-12. ISSN: 2279-0853, ISBN: 2279-0861.
- Bhat S, Sharma R and Euphemia Z (2013).** Carbapenem resistance in clinically significant non fermenting gram negative bacilli. *Journal of Evolution of Medical and Dental Sciences* **2**(47) 9131-9134.

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Clinical and laboratory standard institute (2012). *Performance Standards for Antimicrobial Disc Susceptibility Tests, Twentieth Supplement* 32(3) and M100-S21.

Collee JG, Fraser AG, Marmion BP and Simmons A (1999). *Mackie and McCartney Practical Medical Microbiology*, 14th edition (Churchill Livingstone, Edinburgh (UK)).

Cornaglia G et al., (2007). Study group for antimicrobial surveillance (ESGARS). Metallo-beta-lactamases as emerging resistant determinants in gram negative pathogens: open issues. *International Journal of Antimicrobial Agents* 29 380-8.

Franklin C, Liolios L and Peleg AY (2006). Phenotypic detection of carbapenem- susceptible metallo- β -lactamase- producing Gram-negative bacilli in the clinical laboratory. *Journal of Clinical Microbiology* 44 3139-44

Galani I, Rekatsina PD, Hatzaki D, Plachouras D, Souli M and Giamarellou H (2008). Evaluation of different laboratory tests for the detection of metallo- β -lactamase production in Enterobacteriaceae. *Journal of Antimicrobial Chemotherapy* 61 548-53.

Gupta V, Chhina D And Kaur A (2013). Incidence Of Metallo-B-Lactamase (Mbl) Producing Nonfermenters Isolated From Respiratory Samples In ICU Patients. *International Journal of Pharmaceutical Research and Bio-Science* 4(2) (B) 580 – 585

Hemalatha N, Uma Sekar and Kamat V (2005). Detection of metallo β lactamase producing *Pseudomonas aeruginosa* in hospitalised patient. *Indian Journal of Medical Research* 122 148-52.

Hodiwala A, Dhoke R and Urhekar AD (2013). Incidence of Metallo-Beta-Lactamase Producing *Pseudomonas*, *Acinetobacter* and Enterobacterial Isolates in Hospitalised Patients. *International Journal of Pharmacy and Biological Sciences* 3(1) 79-83

Irfan S, Zafar A, Guhar D, Ahsan T and Hasan R (2008). Metallo- β -lactamase-producing clinical isolates of *Acinetobacter species* and *Pseudomonas aeruginosa* from intensive care unit patients of a tertiary care hospital. *Indian Journal of Medical Microbiology* 26 (3) 243–245.

Jain P, Gandhi V, Patel K, Modi G, Parmar R, Soni S and Vegad MM (2012). Phenotypic Detection Of Metallo-B-Lactamase Producing Enterobacteriaceae. *International Journal of Microbiology Research* 4 (9) 326-329 ISSN: 0975-5276 and E-ISSN: 0975-9174.

Jesudason MV, Kandathil AV and Balaji V (2005). Comparison of two methods to detect carbapenemase and metallo- β -lactamase production in clinical isolates. *Indian Journal of Medical Research* 121 780-783

Kaleem F, Usman J, Hassan A and Khan A. (2010). Frequency and susceptibility pattern of metallo-beta-lactamase producers in a hospital in Pakistan. *The Journal of Infection in Developing Countries* 4(12) 810-813.

Khosravi Y, Loke MF, Chua EG, Tay ST and Vadivelu J (2012). Phenotypic Detection of Metallo- β -Lactamase in Imipenem-Resistant *Pseudomonas aeruginosa*. *The Scientific World Journal* Article ID 654939.

Lee K, Lim YS, Yong D, Yum JH and Chong Y (2003). Evaluation of the Hodge test and the imipenem-EDTA double-disk synergy test for differentiating metallo- β -lactamase producing isolates of *Pseudomonas* spp. and *Acinetobacter* spp. *Journal of Clinical Microbiology* 41 4623-9.

Manikal VM, Landman D, Saurina G, Oydna E, Lal H and Quale J (2000). Endemic carbapenem-resistant *Acinetobacter* species in Brooklyn, New York: Citywide prevalence, inter-institutional spread and relation to antibiotic usage. *Clinical Infectious Diseases* 31 101-6.

Marra AR, Pereira CA, Gales AC, Menezes LC, Cal RG, de Souza JM et al., (2006). Blood stream infections with metallo-beta-lactamase producing *Pseudomonas aeruginosa*: Epidemiology, microbiology and clinical outcomes. *Antimicrob Agents Chemother* 50 388-90.

MHM Nazmul, Jamal H and Fazlul MKK (2012). *Acinetobacter* species-associated infections and their antibiotic susceptibility profiles in Malaysia. *Biomedical Research-India* 23(4): 571-575.

Noyal MJC, Menezes GA, Harish BN, Sujatha S and Parija SC (2009). Simple screening tests for detection of carbapenemases in clinical isolates of nonfermentative Gram-negative bacteria *Indian Journal of Medical Research* 129 707-712.

Research Article

Pandya NP, Prajapati SP, Mehta SJ, Kikani K and Joshi PJ (2011) Evaluation of various methods for detection of metallo- β -lactamase (MBL) production in gram negative bacilli. *International Journal of Biological and Medical Research* **2**(3) 775-777.

Peleg AY, Franklin C, Bell JM and Spelmann DW (2005). Dissemination of the metallo-beta-lactamase gene blaIMP-4 among gram-negative pathogens in a clinical setting in Australia. *Clinical Infectious Diseases* **41** 1549-56.

Pitout JDD, Chow PL, Gregson DB, Laupland KB, Elsayed S and Church DL (2007). Molecular epidemiology of metallo- β -lactamase-producing *Pseudomonas aeruginosa* in the Calgary Health Region: emergence of VIM-2-producing isolates. *Journal of Clinical Microbiology* **45** (2) 294–298.

Tellis R, Muralidharan S and Peter A I (2013). Evaluation of three phenotypic methods for the detection of metallo-beta lactamase production in non fermenting gram negative bacilli. *International Journal of Biomedical and Advance Research* **4**(05).

Varaiya A, Kulkarni M, Bhalekar P, Dogra J. (2008). Incidence of carbapenemresistant *Pseudomonas aeruginosa* in diabetes and cancer patients. *Indian Journal of Medical Microbiology* **26** 238-40.

Walsh TR et al., (2005) Metallo-beta-lactamses: The quiet before the storm? *Clin Microbial Rev* ; **18**; p306-25.

Yong D, Lee K, Yum JH, Shin HB, Rossolini GM and Chong Y (2002). Imipenem-EDTA disk method for differentiation of metallo- β -lactamase-producing clinical isolates of *Pseudomonas* spp. and *Acinetobacter* spp. *Journal of Clinical Microbiology* **40** 3798-801.