

BIOLOGICAL CONTROL OF POWDERY MILDEW DISEASE *SPHAEROTHECA FLUIGINEA* OF *CUCURBITA MAXIMA* (PUMPKIN) SURFACE OF LEAF ANTAGONISTS

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ABSTRACT

Cucurbita maxima (Pumpkin) are cultivated crop members of family cucurbitaceae. The Bottle gourd in the field is affected by several pathogens like bacteria, virus, mycoplasma, nematodes and fungi. Out of 20 surface of leaf isolates of fungi or bacteria. Seven viz. *Trichoderma viride*, *Ampelomyces quisqualis*, *Cladosporium cladosporioides*, *Fusarium pallidoroseum*, *Aspergillus niger*, *Alternaria alternata*, and *Pestalotiopsis dissiminata*. However identified and found antagonistic to *Sphaerotheca fuliginea* (Schlecht ex. Fr.) Pollacci in dual culture on detached leaves of Bottle gourd. *Ampelomyces quisqualis* showed the mycoparasitic activity in dual culture. On the study of antagonism test in vitro. *Fusarium pallidoroseum*, *Cladosporium cladosporioides*, *Pestalotiopsis dissiminata*, and *Ampelomyces quisqualis* were adjudged as potential antagonists of *Sphaerotheca fuliginea*. All the antagonists significantly controlled powdery mildew over check as pre-and post. Inoculation sprays of spore suspension of antagonists on potted plants in glass house. Disease control was better in pre-inoculation spray application of biological control agent as compared to post inoculation application.

Keywords: *Cucurbita Maxima* (Pumpkin), Biocontrol, Fungi

INTRODUCTION

Cucurbita maxima (Pumpkin) is Rabi and Kharif season crop grown all over India in the tropical and sub tropical region. The crop field is attacked by a number of bacterial, fungal viral diseases. Kapoor (1967); Shooty (1971); Goswami and Dasgupta (1981) and Mital *et al.*, (1985) observed powdery mildews on it mainly infected due to *Sphaerotheca fuliginea*. The disease appears in epiphytotic proportions every year and causes considerable losses. None of the management methods provide adequate level of disease control because of sulphur, Bavaistine and Calixin injury to cucurbits, and chemical hazards and non availability of powdery mildew resistant variety (Moursi and Sirry, 1956).

In the literature cited very little information was available regarding the host range of the pathogen; bio chemical changes occurs due to the pathogen and the eco-friendly management of the disease. The study was undertaken with an aim to isolate surface of leaf mycoflora, test their antagonistic activity and control of powdery mildew by potential antagonists in glass house.

MATERIALS AND METHODS

Isolation of surface of leaf bacteria, yeast and fungi were taken from powdery mildew infected and healthy sponge gourd plants by leaf wash method on potato dextrose, malt extract and nutrient broth agar in petriplates. Fungal cultures were purified by hyphal-tip method and single spore isolations. Cultures of bacteria were maintained on PDA and nutrient broth agar slants respectively, under refrigerated conditions. Culture of *Sphaerotheca fuliginea* was maintained on *Cucurbita maxima* plants. Antagonism in dual spore suspension was studied by putting 0.2 ml spore suspension (20 to 25 spores per microscopic field 40x). Each of an antagonists and *sphaerotheca fuliginea* in cavity slides and incubated in humid chamber at 28°C for 24 h. Conidial germination and germ tube length were recorded. Similarly, antagonism on detached leaf was studied by dual culture technique. Inoculated Bottle gourd leaves place in sterilized petriplates containing 200 ppm benzimidazole solution. The distance between was kept at 5 mm. Each treatment was replicated thrice. Colony diameter of both antagonists and *S. fuliginea* was measured 120 h after incubation at 25±1°C percent inhibition in growth of test pathogen over control was

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calculated (Vincent, 1947). Effect of pre-and-post-inoculations with antagonists on powdery mildew development was studied by spraying spore suspension ($5.44 \times 10^6 \text{ ml}^{-1}$) with the help of atomizer on detached leaves of bottle gourd 12 h before and after inoculation and *S. fuliginea*. The inoculated leaves without antagonist served as check. Inoculated leaves in petriplates containing benzimidazole solution were inoculated at $25 \pm 1^\circ\text{C}$ for one week and data on powdery mildew development was recorded. Culture filtrates were obtained by growing antagonist in 250 ml Erlenmayer flask, each containing 50 ml of potato dextrose broth. The flask was inoculated at $25 \pm 1^\circ\text{C}$ for 10 days. Spore suspension of *S. fuliginea* (20 to 25 spores) was prepared in culture filtrates of each antagonists and dispensed (0.2 ml) on clean cavity slides. The slides were incubated in humid chamber at $28 \pm 1^\circ\text{C}$. Data on germination was recorded 72 h after inoculation. In another experiment, culture filtrates were sprayed separately with the help of atomizer on detached sponge gourd leaves in petriplates.

At the same time fresh conidia *S. fuliginea* were dusted on leaves with the help of camel hair brush incubated at $25 \pm 1^\circ\text{C}$ for one week and data on powdery mildew development was recorded. In the glass house experiment, spore suspension ($5.44 \times 10^6 \text{ spores ml}^{-1}$) of promising antagonists were sprayed separately with the help of atomizer 12 h before and after inoculation of *S. fuliginea* on one month old potted plants of bottle gourd (CV. Poinsettle). Four plants were kept in each pot and each treatment was replicated four times. Control plants were maintained without spray of any antagonist. Both the sets of plants were kept in a glass house ($29 \pm 3^\circ\text{C}$) for one month. Data on powdery mildew development was recorded by using 0-4 scale and percent disease index (PDI) was calculated (Mickinney, 1923). The experiment was repeated thrice.

RESULTS AND DISCUSSION

Table 1: Effect of antagonists and its cultural filtrates on conidial germination and germ tube length of *S. fuliginea*

Antagonist	Direct antagonism			Cultural titrate	
	conidial germination %	Inhibition %	Germtube length (μm)	Germination %	Inhibition
<i>Alternaria alternata</i>	14.18	75.00	28.10	5.06	78.10
<i>Cladosporium cladosporioides</i>	15.00	73.76	32.73	11.50	61.38
<i>Fusarium pallidoroseum</i>	08.18	75.16	17.19	008.87	70.34
<i>Aspergillur niger</i>	47.00	23.60	29.00	07.33	81.53
<i>Trichoderma viride</i>	53.13	13.80	29.10	14.80	61.35
<i>Pestalotiopsis dissiminata</i>	22.15	62.47	23.17	12.67	58.21
<i>Ampelomyces quisqualis</i>	08.45	87.18	18.33	04.23	78.87
Check	62.55	---	52.10	34.00	--
LSD (P-0.05)	02.38	04.88	11.50	04.17	9.45

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Table 2: Inhibition of *Sphaerotheca fuliginea* by antagonists and their cultural filtrates on detached leaves of sponge gourd

Antagonist	Direct antagonism			Cultural titrate	
	Pathogen	Antagonist	Inhibition	PDI	Disease control (%)
<i>Alternaria alternata</i>	3.43	2.43	88.97	30.00	82.80
<i>Cladosporium cladosporioides</i>	3.77	2.00	85.70	26.43	87.87
<i>Fusarium pallidoroseum</i>	0.00	4.43	100.00	07.00	92.00
<i>Aspergillus niger</i>	9.43	4.40	35.35	54.00	28.95
<i>Trichoderma viride</i>	5.00	2.40	73.84	71.77	12.10
<i>Pestalotiopsis dissiminata</i>	0.00	5.00	99.80	12.00	91.10
<i>Ampelomyces quisqualis</i>	0.00	4.00	100.00	8.00	99.80
Check	12.00	--	---	83.77	--
LSD (P-0.05)	2.05	2.99	13.47	3.10	4.22

Figures were sine transformed prior to analysis PDF percent disease index.

Table 3: Inhibition of *Sphaerotheca fuliginea* by pre-and-post-inoculation sprays of spore suspension of different antagonists on detached leaf

Antagonist	Pre-Inoculation Spray			Post-Inoculation Spray		
	LFA (%)	LFA(%)	Inhibition	LFA (%)	LFA(%)	Inhibition
	Antagonist	Pathogen	%	Antagonist	Pathogen	%
<i>Alternaria alternata</i>	03.00	15.00	75.89	00.00	14.67	75.09
<i>Cladosporium cladosporioides</i>	00.00	11.00	83.03	12.33	13.33	79.19
<i>Fusarium pallidoroseum</i>	92.00	00.00	98.05	32.67	00.00	100
<i>Aspergillus niger</i>	05.00	52.00	32.23	02.67	54.00	25.97
<i>Trichoderma viride</i>	00.00	28.33	62.88	06.00	22.00	68.36
<i>Pestalotiopsis dissiminata</i>	30.00	00.00	100	06.00	8.77	100
<i>Ampelomyces quisqualis</i>	09.38	00.00	100	03.67	9.43	87.35
Check	---	72.67	---	---	78.33	--
LSD (P-0.05)	07.65	2.22	5.35	13.10	4.32	5.69

LFA = Leaf are covered; figures were are sine transformed prior to analysis.

Table 4: Effect of pre-and post-inoculation spray of potential antagonists on sponge gourd powdery mildew development under glass house conditions

Antagonist	Pre-inoculation spray		Post-inoculation spray	
	PDI	Disease control %	PDI	Disease control %
<i>Fusarium pallidoroseum</i>	28.05	68.00	42.85	48.47
<i>Cladosporium cladosporioides</i>	53.89	29.14	59.00	55.79
<i>Pestalotiopsis dissiminata</i>	74.90	09.46	74.18	10.32
<i>Ampelomyces quisqualis</i>	15.75	89.38	18.50	80.30
Check	82.77	---	82.68	---
LSD (P-0.05)	6.50	8.20	08.50	11.45

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Figures were sine transformed prior to analysis.

To the 20 surface of leaf isolates of fungi and bacteria seven fungi antagonist namely; *Alternaria alternata*, *Aspergillus niger*, *Ampelomyces quisqualis*, *Cladosporium cladosporioides*, *Fusarium pallidoroseum*, *Trichoderma viride* and *Pestalotiopsis dissiminata* were identified and found to be antagonists of *Sphaerotheca fuliginea*. Most of these antagonists have been reported by earlier workers (El-Hammaldehy *et al.*, 1977; Besada, 1978; Srivastava and Bisht, 1986; Szejnberg *et al.*, 1989; Falk *et al.*, 1996; Abo-Foul *et al.*, 1996; Gu and Ko, 1997).

It was observed (Table 1) the results that all the antagonists inhibited conidial germination and reduced germ tube length *Sphaerotheca fuliginea* in dual spore suspension. Maximum conidial inhibition was obtained with *Ampelomyces quisqualis* (87%) followed by *Fusarium pallidoroseum*, *Fusarium pallidoroseum* and *Ampelomyces quisqualis* are resulted in maximum reductions in germ tube length. Barner (1971); Fokkema and Lorbeer (1974); Yadva and Tripathi (1991) and Howell (2003) reported that inhibition of germ tube by antagonists had direct germination of several pathogenic fungi by saprophytic microflora.

The culture filtrate of antagonists also inhibited conidial germination of *S. fuliginea* on cavity slides. Maximum inhibition was obtained with *Ampelomyces quisqualis* *Fusarium Pallidoroseum* and *Pestalotiposis dissiminata*. Similarly, spray of culture filtrate of *Ampelomyces* on detached sponge gourd leaves of sponge gourd resulted 100 per cent control of powdery mildew. However, culture filtrate of *Fusarium pallidoreoseum* as 92 percent disease control respectively. In dual culture on detached bottle gourd leaves, *Fusarium pallidoroseum*, *Pestalotiopsis dissiminata* and *Ampelomyces quisqualis* resulted complete inhibition of *Sphaerotheca fuliginea* (Table 2).

Hashioka and Nakai (1980) reported that the mycoparasitism of *A. quisqualis* on *Erysiphe choracearum*, *Sphaerotheca fuliginea* and other powery mildew causing fungi. Complete inhibition of *S. fuliginea* in present study may be due to mycoparasitism competition for space, nutrient and antibiosis Pre-and Post-inoculation spray of spore suspension of *F. pallidoroseum* and *Pestalotiopsis dissiminata* resulted cent percent control of powdery mildew on detached leaves of sponge gourd (Table 3).

Puzanova (1984) and Dhanbir (2004) reported significant reduction in powdery mildew of cucumber by post inoculation of *Ampelomyces arteminsiae* *A. gypsophitae* *A. gypsophilae* and *A. novvae* *Ampelomyces gisqualis*, *Trichoderma viride*, *Aspergillius fumigantus* in present study showed myloparasitism in dual culture.

It is clear from the (Table 4) that pre-and post-inoculations of all test antagonists gave significant control of sponge gourd powdery mildew on potted plants in glass house. However pre inoculation spray with *Ampelomyces quisaqualis* gave maximum disease control (80.30%). The next best was *Fusarium pallidoroseum* (68%). In general pre inoculation spray was comparatively more effective than post inoculation spray with antagonists. *Cladosporium cladosporioides* as post inoculation spray resulted 56% disease control. Stenijnberg (1979) was reported successful control of *Sphaerotheca fuliginea* on cucumber of *A quisaualis* post inoculation spray of *A. quisqualis* in present investigation resulted 80% disease control.

REFERENCES

- Barner G (1971).** *Ecology of Leaf Surface Micro-organism* (Academic Press) London 581-596.
- Besada WH (1978).** Interaction in plant infect with virus and fungi I. Interaction between each of cucumber mosaic virus and watermelon Mosaic virus against *Sphaerotheca fuliginea* (Schlecht. ex Fr.) Pollaci in some cucurbitace our Hosta. *Acta Phytopathologica Academiae Scientiarum Hungaricae* **13**(1-2) 95-105.
- Dah Dhanbir Sing (2004).** Biocontrol of powdery mildew (*Sphaprothecia fuliginea*) of Cucumber by phylloplane Antagonists. *Journal of Mycology and Plant Pathology* **34**(3) 895-899.
- El-Hammady M, El-Antalby HM and Haroun SH (1977).** Interaction between fungal disease and virus. 1. Powdery mildew and virus infection in cucumber. In: *Current Topics in Plant Pathology*, edited by Kiraly Z (Akademiai Kiado Budapest) 203-208.

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- Falk SPDM, Gadoury P, Cortasi RC Pearson and Secem RC (1995).** Parasitism of *Uncinula necator* Clestothecia by myloparasite. *Ampelomyces quisqualis*. *Phytopathology* **85** 794-800.
- Fokkema NJ and Lorbeer JW (1924).** Interaction between *Alternaria porri* and the saprophytic mycoflora on onion leaves. *Phytopathology* **64** 1128-1133.
- Foul OSVI, Ruskin A Szejnberg and Marder JB (1996).** Disruption of Chlorophyll organization and function in powdery mildew diseased cucumber leaves and its control hyperparasite *Ampelomyces quisqualis*. *Phytopathology* **86** 195-199.
- Goswami BR and Dasgupta MR (1981).** Light blight powdery mildew and charcoal rot disease of sunflower from West Bengal. *Indian Phytopathology* **34** 14-16.
- Gu YM and Ko WH (1997).** Water agarose medium for studying factors affecting germination of *Ampelomyces quisqualis*. *Mycological Research* **101** 422-424.
- Hashioka Y and Nakai Y (1980).** Ultra structure of pycnidial development and mycoparasitism of *Ampelomyces quisqualis* parasite on Erysiphales. *Transactions of the Mycological Society of Japan* **21** 329-338.
- Howell (2003).** Mechanisms employed by *Trichoderma spp.* in the biological control of plant diseases. The history of evolutions of current concepts. *Plant Disease* **87**(1) 4-10.
- Jhooty SS (1971).** Over wintering of powder mildew of cucurbits in Punjab. *Indian Phytopathology* **24** 441-445.
- Kapoor JN (1967).** *Sphaerotheca fuliginea*, *Erysiphe cichoracearum*: CMI descriptions of pathogenic fungi and bacteria set, Kew survey England **16** 152-159.
- McKinney HH (1923).** Influences of soil temperature and moisture on infection of wheat seedlings by *Helminthosporium sativum*. *Journal of Agricultural Research* **26** 195-217.
- Mital SSK Sexena and Akram M (1985).** Effect of powdery mildew fungi *Sphaerotheca fuliginea* on development of Root Knot Nematode *Meloidogyne incognita* on *Lagenaria leucantha*. *Journal of the Indian Botanical Society* **64** 332-305.
- Moursi MA and Sirry AR (1956).** The relation of nitroge and dusting with sulphur of the spread of powdery mildew and yeild of summer squash Ann. *Aquatic Sciences*, California **1** 313-317.
- Puzamova LA (1984).** Hyperparasite of the genus *Ampelomyces* Ces. ex Schlecht and the possibility of their use in the biological control of the pathogen of powdery mildew of plants. *Mikologiya – i – Fitopatologia* **18** 333-338.
- Srivastava SL and Bisht S (1986).** Inhibitory effect of some phyllophlane fungi on powdery mildew disease development. *Indian Phytopathology* **39** 83-86.
- Szejnberg A (1979).** Biological control of powdery mildew by *Ampelomyces quisqualis*. *Phytopathology* **60** 1047 (Abstr.).
- Vincent JH (1947).** Distortion of fungal hyphae in the presence of certain inhibitor. *Nature* **15** 850.
- Waraitch KSGP Munshi and Jhooty JS (1975).** Fungicidal control of powdery mildew of bottle gourd. *Indian Phytopathology* **28** 556-557.
- Warren RC (1970).** Interference by common leaf saprophytes with the development of *Phoma betae* lesions on sugar beet leaves. *Annals of Applied Biology* **72** 137-144.
- Yadva JPS and Tripathi NN (1994).** *Indian Phytopath* **52**(3) 321.