

RAPHIDES IN *URGINEA INDICA* KUNTH (LILIACEAE)

P. Prathima Rao, *M.N. Shiva Kameshwari, Nijagunaiah and K.J. Thara Saraswathi

Department of Botany and Biotechnology, Bangalore University, Bangalore

**Author for Correspondence*

ABSTRACT

Micromorphological traits play a significant role both in systematic and functional aspects of plant survival in response to environmental stimuli. Plant structural trait such as raphides play a vital role in protecting plants from herbivore attack, cause discomfort by physical and chemical irritation by dermal contact to soft tissue thus acts as defensive mechanism. These morphological characteristics may have evolved as response to other environmental stimuli. The presence of raphides (Calcium oxalate crystals) in the anther endothecium represents a rare character, reported in the present study which is of systematic significance. In the present study raphides are present in vegetative and reproductive parts of *Urginea indica*. The mild inflammation and irritation caused when the bulb is rubbed on the skin reveals that it takes part both in mechanical and chemical irritation when they come in contact with tender tissue and protect themselves against herbivore attack. Results indicate that raphide bundle size varies considerably within species. These suggest that, Raphides have some potential to be a useful taxonomic tool. Polarized microscope shows variation in the colouration of raphides.

Key Words: *Idioblastic Cells, Liliaceae, Raphides, Urginea*

INTRODUCTION

Urginea indica Kunth. is a perennial bulbous geophyte native to India, Africa and Mediterranean region and in slopes of hills, in sandy grounds (Gentry *et al.*, 1987; Bruneton, 1996; Bellakhdar, 1997). *U. indica* is a winter plant characterized by three phenological stages consisting of inflorescence, leaves and no above ground biomass, leaves first appear after the flowers have wilted in response to first shower during April to May and may remain green till September depending on rainfall and temperature. Some geophytes that flower without bearing leaves in the beginning of April are known as hysteranthous type and some synanthous type bearing leaves and flowers together (Shiva Kameshwari *et al.*, 2010).

Plants defend themselves against attack from herbivores has been the subject of considerable interest over many decades (Herms and Mattson, 1992; Agarwal and Fishbein, 2006). The plants that are able to survive in environments where herbivores are common because of their ability to resist or recover from intense herbivore pressure (Hartley and Jones, 1997). Cell inclusions, especially different types of calcium oxalate crystals represent significant taxonomic characters at various taxonomic levels in flowering plants. Calcium oxalate crystals have been recorded in most plant families; the most commonly described being the needle shaped raphides and aggregate in bundles within plant cells and are associated with heavy metal tolerance (Franceschi and Nakata, 2005) calcium oxalate crystals are known to protect tree bark from attack by boring insects (Hudgins *et al.*, 2003) and to act as a foliar defense against both invertebrate (Korth *et al.*, 2006) and vertebrate herbivores (Ward *et al.*, 1997). Ward *et al.*, (1997) observed more calcium oxalate crystals in leaves of *Pancreatium sickenbergeri* as it is exposed to the highest rates of gazette herbivory. Raphides are also produced in many economically important plants like Palms, Yams, Banana and Pandanus (See review by Crowther in Press).

Raphide crystals are most commonly encountered among monocots (Dahlgren and Clifford, 1982). The development and chemistry of raphides has been subjected to a few detailed, studies and has been reviewed by Franceschi and Horner (1980).

Our primary goal is to trace raphides present in mucilage of bulbs, in leaves, anthers and in roots of *U.indica* as this will help us to evaluate the irritant present in mucilage which plays a significant role in defense mechanism.

Research Article

MATERIALS AND METHODS

Urginea indica populations were collected from various localities of Karnataka and cultivated in the germplasm of Department of Botany, Bangalore University, Bangalore. Light microscopic studies tissue preparations for anther studies were made by fixing young flower buds in 2.5% gluteraldehyde and 2% paraformaldehyde fixed in 1% osmium tetroxide and dehydrating in an ethanol series. Semi thin sections of 1 μ m thickness were taken using reichert juang ultra microtome and stained with 0.5% toluidine blue O. Photomicrographs were taken in cannon camera attached to nikon microscope.

Maceration of roots were also prepared following Jeffrey's method. The material is treated for one day at 30-40 degree centigrade with mixture of 10% chromic acid and 10% nitric acid.

Fresh material sections of the bulbs were prepared and photographed under Polarized Microscope.

RESULTS

Raphides are present in roots bulbs and leaves. *U. indica* Bulbs which grows inside the soil i.e., in darkness generally produces more idioblasts than leaves exposed to light (Figure 1.A). Raphide crystals are initiated very early in plant development. Individual raphides may be formed by calcium oxalate deposition. Raphides idioblast contains bundles of narrow, elongated needle shaped crystals, usually of similar orientation with pointed ends at maturity, one end is abruptly pointed whereas the other tapers to a point or is wedge shaped. There are varying number of crystals in each bundle (10 to 30).

The frequency of Raphides present in endothecium is considerably higher when compared to other reproductive tissues (Figure 1.B).

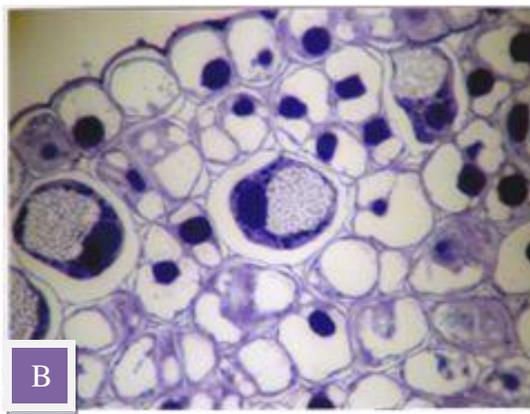
The leaves of *U. indica* are equi-facial i.e., in terms of cuticle, epidermis and spongy cells, which are found on both leaf sides, the lower side of the leaf possesses idioblastic cells containing bundles of raphides.

Raphides occurs within the central vacuole of idioblastic cells in cross section appear foamy and each crystal needle is embedded in a translucent homogeneous substance. In addition, the cell wall of the idioblastic cells contains oil droplets and starch granules (Figure 1.C and D).

The bundles look like a heap of needles in longitudinal section. The length of the raphides vary in different organs (Figure 1.E).

Raphides under polarized Microscope looks like a vibgyor. Each needle like raphides exhibits different colours (Figure 1.F). Bulb sections under the polarized light reveal open bundles of calcium oxalate needles of different sizes. The orientation of Raphide bundles with in a tissue follow the same direction or different direction (Figure 1.G) Sheath walled idioblastic cells containing raphide bundles are present among ordinary cortical cells. Raphides are noticed in both vegetative and reproductive tissues in *U. indica*.

In the macerated root tissues the direction of bundle is parallel to each other (Figure 1.H).



Research Article

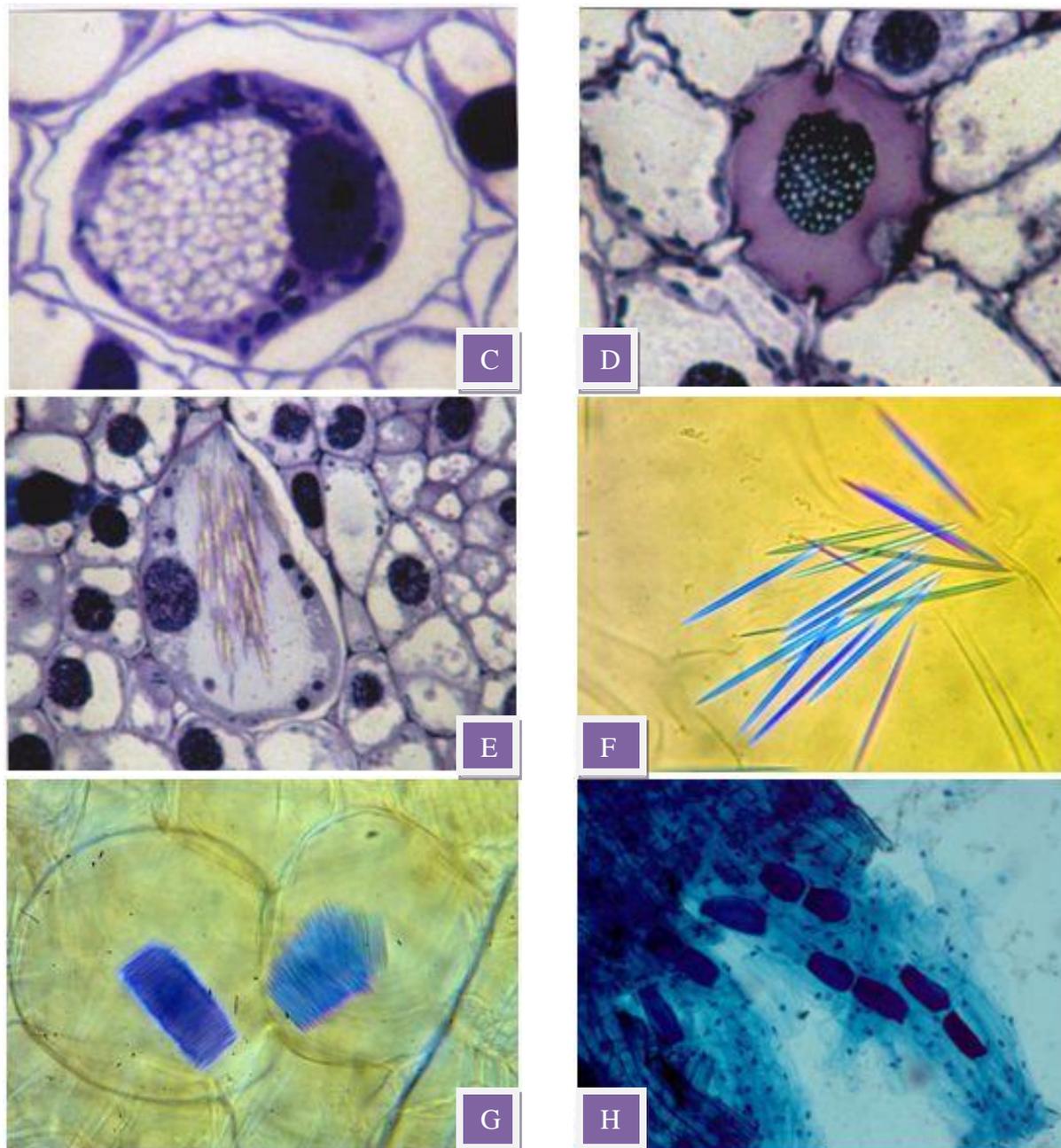


Figure 1: A-*Urginea indica*. B-T.S. of anther section showing Raphides in Endothecium. C-T.S. of Raphide in endothelial cell. D-T.S. of Raphide in middle layers. E-L.S. of Raphide in Endothelial cell. F-Raphide bundles under polarized microscope. G-Bundles of Raphides. H-Bundles of Raphides in Macerated root tissue

DISCUSSION

There is little information of *U. indica* from the biological point of view, except for the Morphological and Karyological studies by (Shiva Kameshwari and Muniyamma, 2004; Professor and Speta, 2004) a few Phytochemical studies (Kopp *et al.*, 1996; Krenn *et al.*, 2000) and some Pharmacological studies

Research Article

(Pascual – Villalobos and Robledo, 1999) It is dominant over wide areas in Tamil Nadu and important for Homeopathic therapy.

Most plants have non cytoplasmic inclusions, such as starch, tannins, silica bodies and calcium oxalate crystals, in some of their cells. Calcium oxalate crystals are widespread in flowering plants, including both dicotyledons and monocotyledons.

They were first discovered by Leeuwenhoek in the 17th Century (Frey, 1929). They have documented using light microscopy (LM) and polarization microscopy and recently using x-ray diffraction infrared and electron Microscopy both scanning (SEM) and transmission (TEM) (Horner and Franceschi, 1981).

The distinct shapes and birefringence, of calcium oxalate crystals, especially raphides and styloids, make them readily observable particularly in young and actively growing plant crystals normally form intracellularly.

The value of calcium oxalate crystals to normal plant growth and development, is largely unknown Frances Chi and Horner (1980s).

They may represent storage forms of calcium and oxalic acid, and there has been evidence of calcium oxalate resorption in times of calcium depletion (Sunell and Healey, 1985) They could also act as simple depositories for metabolic wastes which would otherwise be toxic to the cell or tissue. In some plants they have more specialized functions such as to promote air space formation in aquatic plants or help prevent herbivory. The barbed and grooved raphides of some Aracaceae are particularly irritating to mouth and throat tissues when eaten.

Grooves in crystals which have embedded, themselves in animal tissues may allow the entrance of a chemical irritant such as a toxic proteolytic enzyme or a glucoside into the wound (Sakai, Hanson and Jones, 1972; Walter and Khanna, 1972). Calcium oxalate crystals appear in a variety of shapes which are consistent and repeatable from one generation to the next, demonstrating that the physiological and genetic parameters controlling them are consistent.

Crystals may be present in almost every part of both vegetative and reproductive organs, often in crystal idioblasts near veins, possibly due to calcium being transported through the xylem (Frank, 1967). Crystals are present in leaf epidermal cells in addition to 'normal' bundles of raphides in mesophyll cell (Rudall *et al.*, 1998) Many cellular modifications occur during genesis of crystals, which is a highly complex process (Kausch and Horner, 1983) Raphides are absent in some families of Liliales (Rudall *et al.*, 1996).

In the present study, Endothecium raphides in *U. indica* represents a rare character, such studies have been made by Hardy and Stevenson(2000) in tapetal cells of Commelinaceae and Hamann 1966 in Phylidraceae. Hamann (1966) found that in *Philydrum* almost every tapetal cell contained a raphide bundle, upon degeneration of the tapetum these are released between the developing Pollen grains. As the pollen grains mature crystal quantities decreased and diminished in size. Hardy and Stevenson thus postulated that tapetal crystals had been reabsorbed during pollen grain development, thereby releasing free calcium possibly required in the ontogenetic process. Recently Prychid *et al.*, (2003) observed such cell inclusions in Haemodoraceae.

Raphides appear to occur universally as bundles of needle shaped crystals within vacuolar crystal chambers of idioblastic cells of roots bulbs and leaves. In addition, these crystals also have backward oriented surface bulbs capable of increasing damage to the mouths of grazing animals. Such studies have also been made by Sakai *et al.*, (1972).

Raphides of calcium oxalate in *U.indica* responsible for producing mild inflammation and itching when rubbed on the skin Therefore, raphides take part in both mechanical and chemical irritation when they come into contact with tender tissues of soil-living worms and herbivores. The defense mechanisms could be viewed by other stored compounds to act against Microbial agents, herbivores, rodents, fungi and insect studies made by (Hoffman *et al.*, 1993; Sathyamoorthy *et al.*, 1999; Heth *et al.*, 2000; Civelek and Weintraub, 2004) revealed similar results.

The presence of copious mucilage in *U.indica* is a possible synapomorphy. The end walls of the idioblasts break down as crystal develop, resulting in elongated, asticulate, containing loose groups of raphides

Research Article

often embedded in mucilage. The same results have been given by cogne *et al.*, (2001) in *Urginea altissima*.

The idioblast cells that appeared in *U. indica* is similar to those present in *Allium* species. But one difference is crystals are present in bulb scales of *Allium* but not in aerial leaves according to (Gregory, 1996). But in the present study crystals are present in all parts of the plant body in vegetative and reproductive parts of *Urginea*.

Raphides are present in all organs of *U. indica* examined. It is believed that the highest amount of polysaccharides, accumulates in the underground tissues and this is an adaptive strategy for the plant to survive during dormancy (Sharaf Al-Tar deh *et al.*, 2006). The idioblastic cells contain different phenotypes of crystalloid inclusions. The cell wall contains oil droplets and starch granules.

Initial orientation of calcium oxalate crystals has been improved by using Polarized Microscope and also in Bright Field Light Microscope.

REFERENCES

- Agrawal AA and Fishbein M (2006).** Plant defence syndromes. *Ecology* **87** 132-149.
- Anna H, Lynch, Paula J Rudall and David F Cutler (2006).** Leaf anatomy and systematics of Hyacinthaceae. *Kew bulletin* **61** 145-159.
- Bellakhdar J (1997).** La Pharmacopee Traditionnelle marocaine. Ibis Press, Paris.
- Bruneton J (1996).** Plantes toxiques-vegetaux Pour l'homme Et les animaux Lavoisier, Paris.
- Christin J Pychid and Paula J Rudall (2003).** Systematic significance of cell inclusions in Haemodoraceae and allied families: Silica bodies and Tapetal Raphides. *Annals of botany* **92** 571-580.
- Christina J Prychid, Rachel Schmidt Jabaily and Paula J Rudall (2008).** Cellular ultrastructure and crystal development, in *Amorphophallus* (Araceae). *Annals of Botany* **101** 983-985.
- Christina J Prychid and Paula J Rudall (1999).** Calcium oxalate crystals in Monocotyledons: A review of their structure and systematics. *Annals of Botany* **84** 725-739
- Civelek HS and Wein Traub PG (2004).** Effects of two plant extracts on larval leafminer *Liriomyza trifolii* (Diptera-agromyzidae) in tomatoes. *Journal of Economic Entomology* **97** 1581-1586.
- Cogne AL, Marston A, Mavi S and Hostetmann K (2001).** study of two plants used in traditional medicine used in Zimbabwe for skin problems and rheumatism : *Dioscorea silvatica* and *Urginea altissima*. *Journal of Ethnopharmacology* **75** 51-53.
- Dahlgren RMT and Clifford HT(1982).** The Monocotyledonae: A comparative study. Academic Press, New York.
- Franceschi VR and Makata PA (2005).** Calcium oxalate in plants: Formation and Function. *Annual Review Plant Biology* **56** 41-71.
- Franceschi VR and Horner HT(1980).** Calcium Oxalate Crystals in Plants. *Botanical Review* **46** 361-427.
- Frank E (1967).** Zur building des Kristalli dioblasternmusters bei canavalia eusifformis De. I. *Zeitschirfit fur pflanzen physiologie* **58** 33-48.
- Frey A (1929).** Calcium oxalate Monohydrate and Trihydrat. In: Linsbauer K. Ed. Handbuchder pflanzen anatomie. *Berlin Gebruder Borntraegar* **3** 82-127.
- Gregory M (1996).** Leaf anatomy of *Allium* Sect. *Allium*. In: B. Mathew Ed A review of *Allium* section. *Royal Botanical gardens, Kew* 7-15.
- Gentry HS, Verbisear AJ and Banigan TF (1987).** red squill *Urginea martima* Liliaceae. *Economic botany* **41** 267-282.
- Hamann u (1966).** Embryologische, morphologisch – anatomische and systematische untersuchungen an Philydracean. *Willdenow Bei-heft* **4** 1-178.
- Hardy CR and Stevenson DW (2000).** Development of the gametophytes flower and floral vasculature in *Cachliostema odoratissimum*, (Commelinaceae). *Botanical Journal of the Linnaean Society* **134** 131-157.

Research Article

- Hartley SE and Jones CG (1997).** Plant chemistry and herbivory, or why is the world green ? In: Crawley, M.J. (Ed): Plant Ecology. Blackwell, Oxford 284-324.
- Henry Kraemer (1898).** Origin and detection of Mucilage in Plants. *American journal of pharmacy* **70** 6.
- Herms DA and Mattason WJ (1992).** The dilemma of plants, to grow or defend Q. *Review Biology* **67** 283-335.
- Heth G, Todrank J and Nevo E (2000).** Do spalax Ehrenbergi blind mole rats use food odours in searching for and selecting food ?. *Ethology, Ecology and Evolution* **12** 75-82.
- Hoffman JJ, Timmermann BM, McLaughlin SP and Punna Payak H (1993).** Potential antimicrobial activity of plants from the southwestern united states. *International journal of Pharmacognosy* **31** 101-115.
- Hudgins JW, Krekling T and Francheschi VR (2003).** Distribution of Calcium Oxalate crystals, in the secondary Phloena of conifers constitutive defense mechanism ?. *New Phytologist* **159** 677-690.
- Kausch AP and Horner HT Jr (1983).** The development of Mucilaginous raphide crystal idioblasts in young leaves of Typha angustifolia L. (Typhaceae). *American Journal of Botany* **70** 691-705.
- Kopp B, Krenn L, Draxler M, Hoyer A, Terkola R, Vallaster P and Robien W (1996).** Bufadienolides from *Urginea maritima* from Egypt. *Phytochemistry* **42** 230-233.
- Korth KL, Doege SJ, Park SH, Goggin FL, Wang Q, Gomez SK, Liu G, Jia L and Nakata PA (2006).** *Medicago truncatula* mutants demonstrate the role of plant calcium oxalate crystals as an effective defense against chewing insects. *Plant physiology* **141** 188-195.
- Krenn L, Jelovina M and Kopp B (2000).** New bufadienolides from *Urginea maritima* sensu strictu. *Fitoterapia* **71** 126-129.
- Mick E Hanley, Byron B Lamont, Meredith M Fairbanks and Christine M Rafferty (2007).** Plant structural traits and their role in anti-herbivore defence, perspectives . *plant ecology, evolution and systematics* **8** 157-178.
- Mustopha OT Tour of Biological Research.** Cytotaxonomy of the genus *Urginea* stein, 111. The taxonomic value of foliar anatomical features in *Urginea indica* (Robx). Kunth Complex. *Bioscience research communications* **12**(16) 201-206.
- Nevalainen TJ, Laitio M and Lindgren I (1972).** periodic acid schiff (PAS) staining of Epon-Embedded tissues for light Microscopy. *Acta histochemica* **42** 230-233.
- Pascual Villalobos MJ and Robledo A (1999).** Anti-insect activities of plant extracts from the wild flora in south eastern Spain. *Biochemical systematics and Ecology* **27** 1-10.
- Pfasser MF and Speta F (2004).** From Seilla to Charybdis – is our voyage safer now ? *Plant systematics and Evolution* **246** 245-263.
- Rudall PJ (1998).** Lanariaceae In: Kabitzki K. Ed. The families and genera of vascular plants: III. Flowering plants monocotyledons. Berlin: *Springer verlag* 340-342.
- Rudall PJ, Engelman, EM, Hansen L and Chase MW (1986).** Systematics of Hemiphyllacus, Anemarrhena and Asparagus. *Plant Systematics and Evolution* **211** 181-199.
- Richard C and Keating (2004).** Systematic occurrence of Raphide Crystals in Araceae. *Annals of Missouri Botanical garden* **91**(3) 495-503.
- Sathyamorrthy P, Lugasi-Evgi H, Schlesinger P, Kedar I, Gopas J, Pollack Y and Golan Goldhirsh A (1999).** Screening for cytotoxic and antimalarial activities in desert plants of the Negu and Bedonin market plant products. *Pharma Biology* **37** 188-195.
- Sakai WS, Hamson M and Jones M. (1972).** Raphides with barbs and grooves in *Xanthosoma sangitifolia* Araceae. *Science* **178** 314-315.
- Sharaf Al-Tardeh, Thomas sawidis, Barbara Evelin Diannelidis and Stylianos Delivopoulous (2008).** Morphoanatomical features of the leaves of the Mediterranean geophyte *Urginea maritima* (L) Baker (Liliaceae). *Journal of Plant biology* **51**(2) 150-158.

Research Article

Sharaf Al Tardeh, Thomas sawidis, Barbara Evelin Diannelidis and Stylianos Delivopoulous (2006). Anatomical studies on the adventitious roots of the Geophyte *Urginea maritima*, Baker. *Journal of Biological Research* **75** 61-70.

Shiva Kameshwari MN and Muniyamma M (2004). Karyomorphological studies in populations of *Urginea indica* Kunth Liliaceae. *Beitrage zur Biologie der Pflanzen* **73** 377-394.

Shiva Kameshwari, MN Thara, SaraswathiKJ and Muniyamma M(2005) Morphological variations in populations of *Urginea indica* Kunth Liliaceae. *Journal of Applied and Natural Science* **2(2)** 280-289.

Sultan HAS, Abu Elreish BI and Yagi SM (2010). Anatomical and Phytochemical studies of the leaves and roots of *Urginea grandiflora* Bak and *Pancratium tortuosum* Herbert. *Ethnobotanical leaflets* **14** 826-835.

Sunell IA and Healey PL (1985). Distribution of Calcium Oxalate Crystal idioblasts in leaves of taro *Colocasia esculenta*. *American Journal of Botany* **72** 1854-1860.

Walter WG and Khanna PN (1972). Chemistry of the aroids. I. *Dieffenbachia seguina*, *ameena* and *Pieta*. *Economic botany* **26** 364-372.

Ward D, Spiegel M and Saltz D (1997). Gazelle herbivory and inter population difference in calcium oxalate content of leaves of a desert lily. *Journal Chemistry and Ecology* **23** 333-346.

William C Dickison (2000). Evolution and systematics. Integrative plant anatomy. Academic Press – A Harcourt Science and Technology company, San diego, California USA 205-234.