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AN OVERVIEW OF BIOGENIC SILICA PRODUCTION PATTERN IN THE LEAVES OF *HORDEUM VULGARE* L.

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ABSTRACT

Hordeum vulgare of the family Poaceae is very significant food crop that absorbs biogenic silica in its different parts through their roots and deposited in various cells. The plant cells which were purely silicified after biogenic silica deposition are called as phytoliths. Similarly, present study describes the different type of phytoliths produced by the *Hordeum vulgare* leaf. The most common type of phytoliths found in the leaves of *Hordeum vulgare* are prickly hairs, hair cell phytolith, hair base, rod shaped, epidermal long cell with sinuous margins, sub epidermal long cells, trapezoid, long trapezoid, tracheids, stomata, mesophyll cells and flat epidermal sheet phytoliths. These phytoliths are varied morphologically, morphometrically and also at the level of frequency distributions. These phytoliths provide mechanical support, hardness and strength to plant and also of taxonomically significant.

Keywords: Phytolith, Silica, *Hordeum vulgare*, Poaceae

INTRODUCTION

Phytolith research is an important aid to various disciplines of science like in Plant taxonomy, Plant physiology, Plant molecular biology, Agriculture science, Palaeobotany and Archaeology (Wang and Lu, 1993; Piperno, 2006; Lu *et al.*, 2007; Madella *et al.*, 2009; Chauhan *et al.*, 2011; Tripathi *et al.*, 2011, 2012 a, b, 2013).

Phytoliths are basically plant silica stones made up of silicon dioxide. Among all elements silica is regarded as one of the most beneficial element for the plants that protects them from various biotic and abiotic stresses (Tripathi *et al.*, 2012 a, b, c, 2013, 2014; Ma and Yamaji, 2006). Plants accumulate silica in the form of monosilicic acid, Si (OH)₄ and deposited in and between various types of the plant cells (Tripathi *et al.*, 2011, 2012 a,b,c,d).

It has been observed that comparatively, phytoliths are highly abundant in leaves followed by the inflorescence, leaf sheaths and stem (Chauhan *et al.*, 2011a, b; Tripathi *et al.*, 2012 a, b). Phytolith reference collection of various families and genera is carried out all over the world, however phytolith analysis of Poaceae plants has a core attention aspect among the phytolith researchers while In India it is lacking.

Therefore, in the present study we have selected *Hordeum vulgare* L. belonging to Poaceae family to analyze its phytolith production pattern in general.

Hordeum is a genus consist about 30 species of annual and perennial grasses, inhabitant of the temperate area all over the world. *H. vulgare* species is principally used as food crops for human beings and as a fodder for animals while it also have a major commercial importance (Purseglove, 1974).

The formation and deposition of phytoliths in various cereals including *Hordeum* was investigated earlier by several workers which may include like Blackman (1968, 1969), Blackman and Parry (1968), Hayward and Parry (1973), Hodson and Sangster (1989), Hutton and Norrish (1974), Jones and Handreck (1965), Kaufman *et al.*, (1972), Soni and Parry (1973).

Although the phytoliths of *Hordeum vulgare* inflorescence were described by Ball *et al.*, (1999); Ball *et al.*, (2009); Honain *et al.*, (2006) but no detailed study of *Hordeum vulgare* leaf phytoliths has been taken yet. Therefore, we have described the phytolith types, measurements and frequency distribution of *Hordeum vulgare* leaf in detail.

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MATERIALS AND METHODS

The samples of *Hordeum vulgare* were collected from the Roxburgh Botanical Garden, Department of Botany, University of Allahabad. Leaves of *Hordeum vulgare* were subjected to a carefully washing in distilled water to remove the dust particles from the surface of the samples and were dried at 60°C for 48 hrs. After removing the moisture from the samples, they were positioned in porcelain crucibles and reduced to ash in a muffle furnace at about 400°C to 500°C for 4-6 hours until the ash emerged whitish in color. Consequently the ash was treated with nitric acid (HNO₃) and rest were rinsed with distilled water and centrifuged. The residual biogenic silica was dried and mounted in Canada balsam using 0.001 gm ash/ slide and ten slides were prepared. Difference in frequency distribution of silicified cells as well as their shape and size was observed in ten slides under the area of 1.24 mm²/ slide (Chauhan et al., 2011; Tripathi et al., 2012; 2013).

RESULTS AND DISCUSSION

Studies suggested that different types of cells are present in the plant leaves in which silica is finally deposited likewise these silicified cells are called as phytoliths. Figure 1 depicts the variety of phytoliths present in the leaves of *Hordeum vulgare* i.e. prickle hairs (1-9), hair cell phytolith (10), hair base (11), rod shaped (12,13), epidermal long cell with sinuous margins (14-17), sub epidermal long cells (18), trapezoid (19), long trapezoid (20), tracheids (21), stomata (22) mesophyll cells and flat epidermal sheet. These phytoliths are taxonomically very important in identifying plants at genus or at family level, beside this, silicified structures also play an important role to protect the plants in various ways, such as they affected the optical properties of the leaves after interaction with sun light and also improve the strength of plants (Goto *et al.*, 2009; Klancnik *et al.*, 2014). Klancnik *et al.*, (2014) suggested the considerable roles of the various silicified leaf structures found at near-surface area such as prickle hairs, cuticle and epidermis on the basis of redundancy investigations. Therefore, it may assume that phytoliths present in the leaves of *Hordeum vulgare* perform various roles to protect the plants in adverse environmental conditions. Another role of silicified cells in carbon sequestration is a new area of research which interestingly carrying out by phytolith researchers in all over the world. Carbon entrapped by the silicified cells is extremely resistant and decomposed in the soil for several thousands of years after plant decay which clearly reveal the significance of silicified cells (phytoliths) in the continuing biogeochemical sequestration of atmospheric CO₂ (Song *et al.*, 2013).

Stomata are very important internal machinery of plants by which plants performs various metabolic and physiological responsibilities to protect the plants as well as regulate the essential phenomena. It has also been reported that various silicified cells along with stomata may be a good indicator to detect the level of CO₂ in the paleo-atmosphere (Parr and Sullivan 2011; Song *et al.*, 2013). Similarly figure 1 of this study shows silicified stomata found in the leaves of *Hordeum vulgare* thus phytolith analysis of this study may also be helpful to detect the level of CO₂ in the paleo-atmosphere, if *Hordeum vulgare* leaf phytoliths were suspected in the soil samples of archaeological sites.

It has been recommended after the morphological observations of phytoliths that, their morphometry have also very important contributions in phytolith research because various studies revealed that morphology of different types of phytoliths in different plant species or genera were more or less similar however, size of these phytoliths may varied. Thus morphometrical study of each phytolith type may helps to discriminate the plants at different level (Tripathi *et al.*, 2011, 2012a, b; Madella *et al.*, 2009; Ball *et al.*, 1999; Ball *et al.*, 2009; Honain *et al.*, 2006). In this study we have also measured the size of each phytolith which are different among each other due to differences in their particular shapes (Table1). Measurements of phytoliths shows that long epidermal phytoliths are highest in length followed by the hair cell phytoliths, sub epidermal or rod-shaped, large-prickle phytoliths, stomata, long trapezoids, short epidermal phytoliths, small prickle phytoliths and short trapezoids. Beside stomata of the *Hordeum vulgare* leaf shows highest in width followed by the large-prickle phytoliths, small prickle phytoliths, long epidermal phytoliths, sub epidermal or rod-shaped, short epidermal phytoliths, hair cell phytoliths,

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long trapezoids and short trapezoids. These observations clearly depict the variations among the phytoliths of *Hordeum vulgare* that is because of variation in the original size of plant cells.

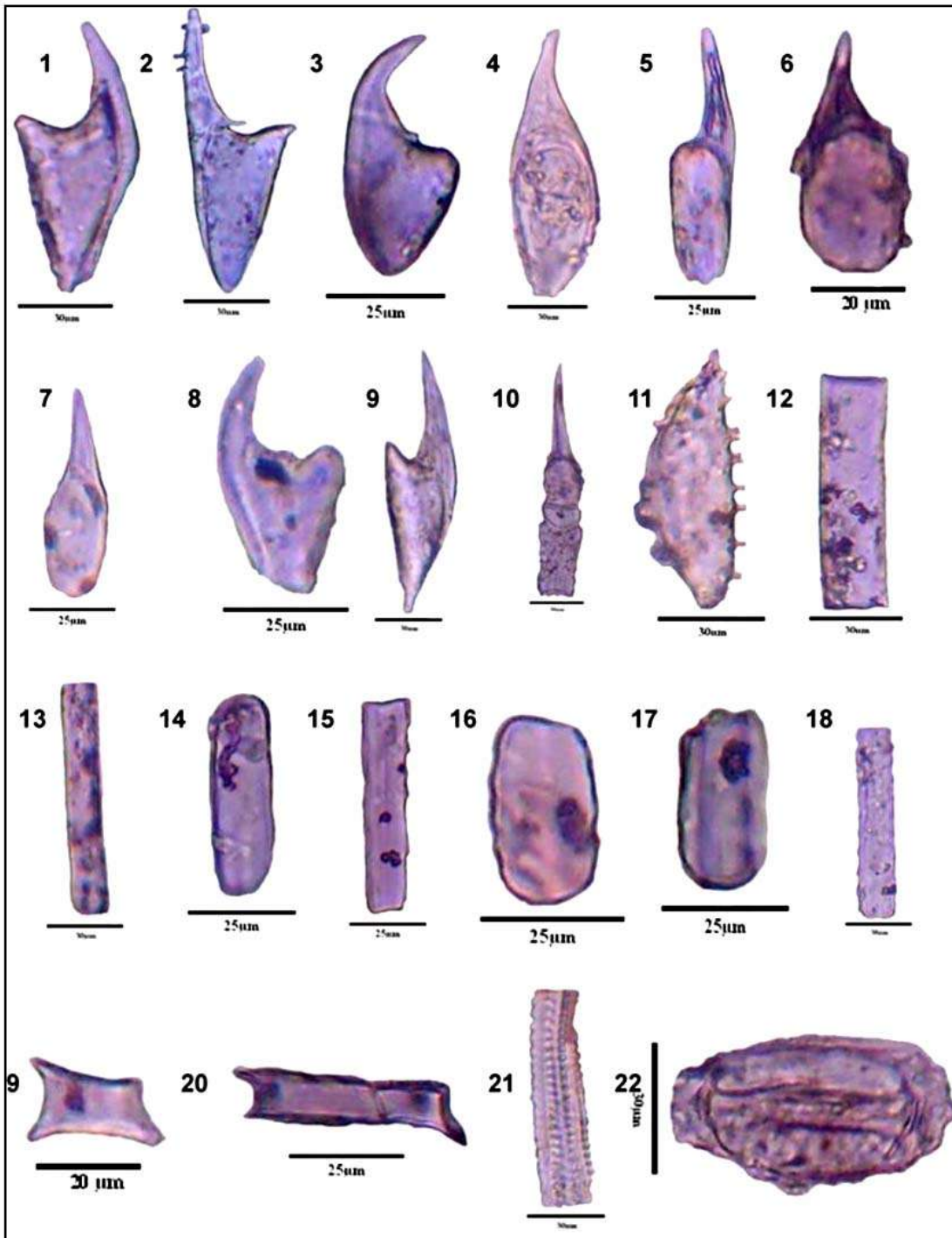


Figure 1: Different type of leaf phytoliths of *Hordeum vulgare*: prickle hairs (1-10), hair cell phytolith (10), hair base (11), rod shaped (12, 13), epidermal long cell with sinuous margins (14-17), sub epidermal long cells (18), trapezoid (19), long trapezoid (20), tracheids (21), stomata (22).

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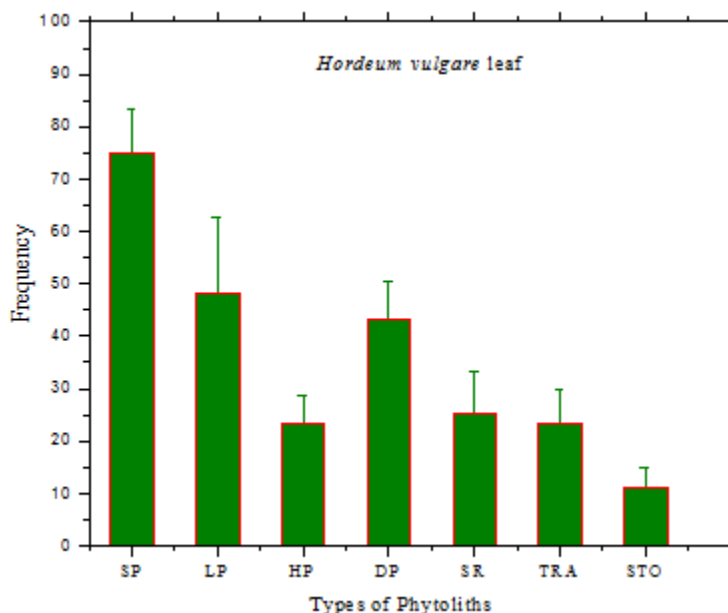


Figure 2: Frequency of different types of phytoliths of *Hordeum vulgare* leaves: small prickle phytoliths (SP), large-prickle phytoliths (LP), hair cell phytoliths (HP), long epidermal phytoliths with sinuous margins (DP), short epidermal phytoliths (SR), trapezoids (TRA), stomata (STO).

Table 1: Measurements of different phytoliths of *Hordeum vulgare* leaves

Phytolith Types	Length(µm)	Width(µm)
Small prickle phytoliths	35.6± 11.43	16.87± 2.39
Large-prickle phytoliths,	75.5± 15.2	19.37±4.26
Hair cell phytoliths,	118.75±19.4	10±3.53
Long epidermal Phytoliths	200±25	17.5±2.5
Short epidermal Phytoliths	44.16± 6.29	13.12±4.2
Sub epidermal or rod-shaped	82.5± 13.9	13.3±3.81
Long trapezoids	48.33±10.10	10±2.5
short trapezoids	16.25±3.2	7.5±2.5
Stomata	50.83±6.29	24.3±6.02

Furthermore, in recent years of phytoliths research, some issues of phytolith quantification i.e. phytolith counts are important or not for discrimination of plants and its species are gaining much interest between the phytoliths researcher and also have a matters of interesting debates (Strömberg, 2009a, b; Alexandre and Bremond, 2009). In this study quantification of *Hordeum vulgare* phytoliths were also performed which deals highest frequency of short prickle phytoliths followed by the long prickle, hair cell, epidermal long cell phytolith with sinuous margins, short epidermal rod shaped, trapezoid and stomata (figure 2). Chauhan *et al.*, (2011) and Tripathi *et al.*, (2012) successfully reported the variation of frequency distribution in each type of phytoliths in *Arundo donax* and two species of *Sorghum*.

Previous studies of *Hordeum vulgare* were mainly focused on their inflorescence phytolith which did not address combined issues like morphology, morphometry and frequency of phytoliths and also does not

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discussed the leaf phytolith production pattern (Blackman, 1969; Hutton and Norris, 1974; Hayward and Parry, 1973; Bennett, 1982; Hodson and Sangster, 1988, 1989; Ball *et al.*, 1993). This study documented the phytolith production pattern of *Hordeum vulgare* leaf along with their morphology, morphometry and frequency distribution. Further this study may be helpful to resolve some physiological, archaeobotanical and taxonomical issues, however detailed study of other parts of *Hordeum vulgare* as well as comparison among the *Hordeum* species are needed.

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REFERENCES

- Alexandre A and Bremond L (2009).** Comment on the paper in Quaternary International: Methodological concerns for analysis of phytolith assemblages: Does count size matter? (CAE Strömberg), *Quaternary International Journal* **193**(1) 141-142.
- Ball TB, Brotherson JD and Gardner JS (1993).** A typologic and morphometric study of variation in phytoliths from inkhorn wheat (*Triticum monococcum* L.). *Canadian Journal of Botany* **71** 1182-1192.
- Ball TB, Ehlers R and Standing MD (2009).** Review of typologic and morphometric analysis of phytoliths produced by wheat and barley. *Breeding Science* **59** 505-512.
- Ball TB, Gardner JS and Anderson N (1999).** Identifying inflorescence phytoliths from selected species of wheat (*Triticum monococcum*, *T. dicoccon*, *T. dicoccoides*, and *T. aestivum*) and barley (*Hordeum vulgare* and *H. spontaneum*). *American Journal of Botany* **86** 1615-1623.
- Bennett DM (1982).** Silicon deposition in the roots of *Hordeum sativum*, *Avena sativa* L. and *Triticum aestivum* L. *Annals of Botany* **50** 239-246.
- Blackman E (1968).** The pattern and sequence of opaline silica deposition in rye (*Secale cereale* L.). *Annals of Botany* **32** 207-218.
- Blackman E (1969).** Observations on the development of the silica cells of the leaf sheath of wheat (*Triticum aestivum*). *Canadian Journal of Botany* **47** 827-838.
- Blackman E and Parry DW (1968).** Opaline silica deposition in rye (*Secale cereale* L.). *Annals of Botany* **32** 199-206.
- Chauhan DK, Tripathi DK, Kumar D and Kumar Y (2011a).** Diversity, distribution and frequency based attributes of phytolith in *Arundo donax* L. *International Journal of Innovation in Biological and Chemical Sciences* **1** 22–27.
- Chauhan DK, Tripathi DK, Rai NK and Rai AK (2011b).** Detection of biogenic silica in leaf blade, leaf sheath, and stem of bermuda grass (*Cynodon dactylon*) using LIBS and phytolith analysis. *Food Biophysics* **6** 416–423.
- De Silva D and Hillis WE (1980).** The contribution of silica to the resistance of wood to marine borers. *Holzforschung* **34** 95-97.
- Purseglove, J W. 1974.** Tropical crops monocotyledons. Vol.1. Longman group. Ltd., London
- Goto M, Hiroshi E, Shyuichi K, Keiji T, Natsumi O, Yutaka Y, Satoru O, Mohammed SY and Osamu M (2003).** Protective effect of silicon on phenolic biosynthesis and ultraviolet spectral stress in rice crop. *Plant Science* **164**(3) 349-356.
- Hayward DM and Parry DW (1973).** Electron-probe microanalysis studies of silica deposition in barley (*Hordeum sativum* L.). *Annals of Botany* **37** 579-591.
- Hodson MJ and Sangster AG (1988).** Observation on the distribution of mineral elements in the leaf of wheat (*Triticum aestivum* L.) with particular reference to silicon. *Annals of Botany* **62** 463-471.

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Hodson MJ and Sangster AG (1989). Silica deposition in the inflorescence bracts of wheat (*Triticum aestivum*) II. X-ray microanalysis and backscattered electron imaging, *Canadian Journal of Botany* **67** 281-287.

Honaine MF, Zucol AF and Osterrieth ML (2006). Phytolith Assemblages and Systematic Associations in Grassland Species of the South-Eastern Pampean Plains, Argentina. *Annals of Botany* **98** 1155-1165.

Hutton JT and Norrish K (1974). Silicon content of wheat husks in relation to water transpired. *Australian Journal of Agricultural Research* **25** 203-212.

Jones LHP and Handreck KA (1965). Studies of Silica in the oat plant, III. Uptake of silica from soils by the plant. *Plant and Soil* **23** 79-96.

Kaufman PB, Soni SL, Lacroix JD, Rosen JJ and Bigelow WC (1972). Electron-probe microanalysis of silicon in the epidermis of rice (*Oryza sativa* L.) internodes. *Planta* **104** 10-17.

Klančnik K, Vogel-Mikuš K and Gaberščik A (2014). Silicified structures affect leaf optical properties in grasses and sedge. *Journal of Photochemistry and Photobiology B: Biology* **130** 1-10.

Lu HY, Wu NQ, Liu KB, Jiang H and Liu TS (2007). Phytoliths as quantitative indicators for the reconstruction of past environmental conditions in China II: palaeoenvironmental reconstruction in the Loess plateau. *Quaternary Science Reviews* **26**(5-6), 759-772.

Ma JF and N Yamaji (2006). Silica uptake and accumulation in higher plants. *Trends in Plant.*

Madella M, Jones MK, Echlin P, Powers-Jones A and Moore M (2009). Plant water availability and analytical microscopy of phytoliths: implications for ancient irrigation in arid zones. *Quaternary International Journal* **193** 32-40.

Parr JF and Sullivan LA (2011). Phytolith occluded carbon and silica variability in wheat cultivars. *Plant and Soil* **342**(1-2) 165-171.

Piperno DR (2006). Phytoliths: a comprehensive guide for archaeologists and paleoecologists. Rowman Altamira.

Singh VP, Tripathi DK, Kumar D and Chauhan DK (2011). Influence of exogenous silicon addition on aluminium tolerance in rice seedlings. *Biological Trace Element Research* **144**(1-3) 1260-1274.

Song Z, Parr JF and Guo F (2013). Potential of Global Cropland Phytolith Carbon Sink from Optimization of Cropping System and Fertilization. *PloS one* **8**(9) e73747, 1-6

Soni SL and Parry DW (1973). Electron probe microanalysis of silicon deposition in the inflorescence bracts of the rice plant. (*Oryza sativa*). *American Journal of Botany* **60** 111-116.

Strömberg CA (2009). Methodological concerns for analysis of phytolith assemblages: Does count size matter?. *Quaternary International Journal* **193**(1) 124-140.

Strömberg CA (2009). Reply to comment on Methodological concerns for analysis of phytolith assemblages: Does count size matter? (A. Alexandre and L. Brémond). *Quaternary International Journal* **193**(1) 143-145.

Tripathi DK, Chauhan DK, Kumar D and Tiwari SP (2012a). Morphology, diversity and frequency based exploration of phytoliths in *Pennisetum typhoides* Rich. *National Academy Science Letters* **35**(4) 285-289.

Tripathi DK, Kumar R, Chauhan DK, Rai AK and Bicanic D (2011). Laser-induced breakdown spectroscopy for the study of the pattern of silicon deposition in leaves of saccharum species. *Instrumentation Science and Technology* **39** 510-521.

Tripathi DK, Kumar R, Pathak AK, Chauhan DK and Rai AK (2012b). Laser-Induced Breakdown Spectroscopy and Phytolith Analysis: An Approach to Study the Deposition and Distribution Pattern of Silicon in Different Parts of Wheat (*Triticum aestivum* L.) Plant. *Agricultural Research* **1**(4) 352-361.

Tripathi DK, Mishra S, Chauhan DK, Tiwari SP and Kumar C (2013). Typological and frequency based study of opaline silica (phytolith) deposition in two common Indian Sorghum L. species. *Proceedings of the National Academy of Sciences, India Section B: Biological Sciences* 1-8.

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Tripathi DK, Singh VP, Gangwar S, Prasad SM, Maurya JN and Chauhan DK (2014). Role of Silicon in Enrichment of Plant Nutrients and Protection from Biotic and Abiotic Stresses. In *Improvement of Crops in the Era of Climatic Changes*, Springer New York 39-56.

Tripathi DK, Singh VP, Kumar D and Chauhan DK (2012c). Rice seedlings under cadmium stress: effect of silicon on growth, cadmium uptake, oxidative stress, antioxidant capacity, and root and leaf structures. *Chemistry and Ecology* **28** 281–291.

Tripathi DK, Singh VP, Kumar D and Chauhan DK (2012d). Impact of exogenous silicon addition on chromium uptake, growth, mineral elements, oxidative stress, antioxidant capacity, and leaf and root structures in rice seedlings exposed to hexavalent chromium. *Acta Physiologiae Plantarum* **34** 279–289.

Wang YJ and Lu HY (1993). *The Study of Phytolith and its Application*. China Ocean Press, Beijing, 228 (in Chinese).